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Article in *Journal of Pest Science* · March 2016

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# Non-crop plants used as hosts by *Drosophila suzukii* in Europe

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Received: 13 November 2015 / Revised: 4 March 2016 / Accepted: 14 March 2016 / Published online: 24 March 2016  
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**Abstract** The invasive spotted wing drosophila *Drosophila suzukii*, a fruit fly of Asian origin, is a major pest of a wide variety of berry and stone fruits in Europe. One of the characteristics of this fly is its wide host range. A better knowledge of its host range outside cultivated areas is essential to develop sustainable integrated pest management strategies. Field surveys were carried out during two years in Italy, the Netherlands and Switzerland. Fruits of 165 potential host plant species were collected, including mostly wild and ornamental plants. Over 24,000 *D. suzukii* adults emerged from 84 plant species belonging to 19 families, 38 of which being non-native. Forty-two plants were reported for the first time as hosts of *D. suzukii*. The highest infestations were found in fruits of the genera *Cornus*, *Prunus*, *Rubus*, *Sambucus* and *Vaccinium* as well as in *Ficus carica*, *Frangula alnus*, *Phytolacca americana* and *Taxus baccata*. Based on these data, management methods are suggested. Ornamental and hedge plants in the vicinity of fruit crops and orchards can be selected according to their susceptibility to *D. suzukii*. However, the widespread availability and abundance of non-crop hosts and the lack of efficient native parasitoids suggest the need for an area-wide control approach.

**Keywords** Spotted wing drosophila · Fruit fly · Host range · Invasive species

## Key message

- *Drosophila suzukii* a fruit fly of Asian origin has a broad host range. Field surveys in Europe identified more than 80 host plants including wild ornamental and crop plants.
- Knowing the host range of *D. suzukii* outside cultivated habitats is essential for the development of sustainable IPM strategies.
- The widespread availability of non-crop hosts and the lack of efficient parasitoids suggests the need for an area-wide control approach.

## Introduction

The spotted wing drosophila, *Drosophila suzukii* Matsumura (Diptera: Drosophilidae), is a fruit fly of East-Asian origin that rapidly invaded other parts of the world in the late 2000s (Cini et al. 2012; Deprá et al. 2014; Asplen et al. 2015). In contrast to most other *Drosophila* spp. that develop only on overripe or decaying fruits, *D. suzukii* is able to oviposit in ripe fruits due to the female's prominent serrated ovipositor (Lee et al. 2011; Walsh et al. 2011). Since its first notification in 2008 in Europe, it has rapidly spread to most suitable areas of the continent (Cini et al. 2014), becoming a major pest of a wide variety of berry and stone fruit crops (Asplen et al. 2015).

Before becoming a worldwide invasive species, *D. suzukii* was considered a relatively minor pest in its area of

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Special Issue: Spotted Wing Drosophila

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Communicated by A. Biondi.

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origin (Asplen et al. 2015; Haye et al. 2016) and, therefore, efficient management techniques were not available. However, research on management methods has been carried out recently in various parts of the world (Haye et al. 2016). *Drosophila suzukii* shows at least three important biological characteristics that may strongly influence the development of integrated management methods. Firstly, *D. suzukii* is poorly attacked by natural enemies in the invasion range (Haye et al. 2016). In particular, larval parasitism is almost never observed, while *Drosophila* spp. larvae are usually heavily parasitised by braconid and figitid wasps (Carton et al. 1986). Laboratory assays suggest that European and American larval parasitoids are not able to develop on *D. suzukii*, apparently because of the strong host immune response of the invasive fly against these parasitoids (Chabert et al. 2012; Kacsoh and Schlenke 2012; Poyet et al. 2013). Secondly, its fast development (ca. two weeks to develop from egg to adult at 22 °C (Tochen et al. 2014) allows it to produce many generations per year between spring and autumn. In the temperate climate of Oregon, Western USA, Tochen et al. (2014) estimated that *D. suzukii* undergoes an average of 7.1 generations per year, but up to 13 generations per year have been cited for warmer climates (Asplen et al. 2015); in temperate climates, the winter is spent as adults in reproductive diapause (Zerulla et al. 2015). Thirdly, *D. suzukii* has a very broad host range, including fruits of many wild and ornamental host plants (Lee et al. 2015; Poyet et al. 2015), which allow it to move regularly from cultivated to wild and urban habitats. These characteristics imply that the potential development of the fly in wild and ornamental fruits in the vicinity of orchards and fruit fields has an important impact on the level of attack in cultivated fruits.

For obvious reasons, the host range of *D. suzukii* among cultivated fruits has been assessed extensively (e.g. Mitsui et al. 2010; Lee et al. 2011; Bellamy et al. 2013; Burrack et al. 2013) whereas less emphasis has been placed on non-cultivated hosts. Very recently, however, wild and ornamental non-crop hosts have been studied in Michigan and Oregon (USA) by Lee et al. (2015) who found *D. suzukii* in 24 field-collected plant species belonging to 12 families. They also made additional assessments of host suitability in laboratory tests and provided a literature review on the host range of the fly worldwide. In Europe, the most extensive host range study is that of Poyet et al. (2015). They tested, in the laboratory, *D. suzukii* on 67 fruit species collected in Northern France and found out that *D. suzukii* laid eggs on 50 of them and successfully developed in 33, belonging to 15 families. However, there have been discrepancies between host range data gathered from field surveys and laboratory tests in North America (Lee et al. 2015) and, so far, no extensive field survey was carried out

to assess the realised host range of *D. suzukii* in non-crop hosts in Europe.

The main objective of this study was to assess the host range realised by *D. suzukii* outside cultivated areas in Western and Central Europe. For this, surveys were carried out during two years in the Netherlands, Northern Italy and Switzerland to collect fruits in semi-natural and urban landscapes and rear out *D. suzukii*. Attempts were made to classify the host fruits according to the frequency and level of infestation.

## Materials and methods

Potential host fruits, i.e. fruits that appeared sufficiently soft to allow the oviposition and development of *D. suzukii*, were collected through regular surveys in 2014 and 2015 at various sites in three countries: Italy, the Netherlands and Switzerland. In all three countries, sites were distant from each other's by at least one km. All sampling regions had been infested by *D. suzukii* at least since 2013, and the presence of *D. suzukii* in the areas during the sampling periods was confirmed by trapping campaigns for monitoring adult populations. The survey focused on ripe fruits of wild and ornamental non-crop hosts in various habitats, i.e. forests, forest edges, meadows, hedges in agricultural habitats, gardens and parks, etc. In a few cases, fruit trees planted as urban or garden trees at the surveyed sites were also sampled. Plants were identified using local reference guides (Pignatti 1982; Meijden 1996; Ferrari and Medici 2008; Koning and Broek 2012; Info Flora 2015). Sampling techniques were rather similar in the three countries but with some differences. Therefore, they are described separately below.

### Italy

Twenty-nine sites in semi-natural habitats located in seven different areas in North-eastern Italy (Veneto and Trentino Regions) were sampled every two weeks from March 2014 to October 2015. The fruits were collected when available from all potential host plants. Moreover, occasional collections were made in various landscapes in Liguria, Toscana and Veneto Region wherever new fruit species were found. Fruit were sampled from a total of 116 plant species. When possible, samples consisted in 2 dl of small fruits or 50 individuals of large fruits, but smaller amount of less abundant fruits were also collected. For each sample, the number of fruits was counted and their weight was measured. Fruits were then stored in containers, covered with fine mesh and kept at 23 °C. Emerging insects were collected three times a week and lasted three weeks after the last emergence of *D. suzukii*. Flies were stored in ethanol for later identification.

## Netherlands

Three areas were selected in the centre of the Netherlands. The areas differed in respect to soil and vegetation type. The first was in the orchard dominated river clay area in Gelderland province. The second was in a semi-urban area in the Utrecht province, where river clay meets the sandy Pleistocene soils. The third was in forests and at forest edges on the sandy Pleistocene soils in Gelderland province. At each area, surveys were made at three sites of 0.5 ha each. The vegetation at each of the nine sites was sampled eight times from June to October 2014. Additionally, a large sampling effort was made on December 4, 2014, to determine whether *D. suzukii* could overwinter as a larva in fruits. In 2015, surveys were carried out between May and October at the same sites. At each sampling date, fruits were picked from all potential host plants. Occasional collections were also made in the region, wherever new potential host plant species were found. Fruits of 34 plant species were collected in 2014 and 68 in 2015. In total, 77 different plant species were sampled. When possible, samples consisted of ca. 50 fruits, but smaller numbers of less abundant fruits, or larger numbers of abundant but small fruits were also collected. After weighing, fruits were put in containers, covered with fine mesh and kept at 22 °C. Emerging insects were collected three times a week until three weeks after the last emergence of *D. suzukii* and stored in ethanol for later identification.

## Switzerland

Collections were carried out only in 2014, mainly in the Canton Ticino, in the Southern Alps. Fruits of a variety of potential host species were collected at ten sites, once per month, from early May to early October 2014. Additional collections were made along elevation gradients in the Ticino, once in July and once in August 2014. Some collections were also made in the Jura Canton, Northwestern Switzerland. A total of 39 plant species were sampled. When possible, samples consisted of ca. 50 fruits, but smaller numbers of less abundant fruits, or larger numbers of abundant but small fruits were also collected. For each sample, the number of collected fruits was recorded. Fruits were then placed in photo-electors made of a cardboard cylinder surrounded by a funnel ending in a translucent plastic cup, in which the flies were collected daily until three weeks after the last emergence of *D. suzukii*. They were then killed in ethanol to allow a careful counting of the number of *D. suzukii* adults. After the emergence period, the cylinders were inspected to count the few flies that had died without reaching the cup.

## Data analyses

Two parameters were calculated: the rate of occurrence and the infestation level. The rate of occurrence expressed the geographical frequency at which *D. suzukii* was found on a particular fruit species, without taking into account the level of attack at specific sites. It was calculated as the ratio between the number of sites × years (throughout all three countries) where a fruit was found attacked by *D. suzukii* divided by all sites × years combinations where the fruit was collected.

To allow a comparison of the infestation level among host species, the number of flies emerging per individual fruit is not a very good parameter because fruit size strongly varies among species. Instead, the number of flies should be expressed per fruit weight, volume or skin surface. In Italy and Switzerland, all fruits were counted but the size and weight of fruits could not be measured for all samples. Thus, for each fruit species collected in Italy and Switzerland, the average diameter was gathered in the literature, mainly in Info Flora (2015) and, if not indicated, an average of the average data found in various information sources (other books on regional flora and web sites from scientific societies and organisations) was calculated. In case of oval fruits, the length and the width were averaged. The fruit surface was estimated for each species ( $\text{surface} = 4\pi r^2$ ). Aggregate fruits composed of drupelets, e.g. *Rubus* spp., were treated in the same way, although we realise that, for these fruits, the surface was underestimated. Then, for each sample, a level of infestation was expressed as the number of *D. suzukii* adults emerged per  $\text{dm}^2$  of fruit surface. In the Netherlands, the number of fruits was not counted but, instead, samples were weighed. Thus, for these samples, the level of infestation was expressed as the number of *D. suzukii* adults per kg of fruit. We realise that none of these two parameters are perfect. The fruit surface is probably a better expression of the potential of the fruit to attract *D. suzukii* and to support the development of a certain quantity of larvae than its weight or its volume. On the other hand, for some species, the size of the sampled fruits may be rather different from the average size found in the literature. Furthermore, if the fruit is very small in size individually, the fruit surface may not matter as much, and having other measures might be useful. But the aim of this parameter was not to finely compare fruit species but rather to broadly categorise the infestation levels of host fruits in the field. For a finer comparison of infestation levels, several confounding factors such as time of collection, habitat, fruit density and population size of the flies would have to be taken into account. For the same reasons, the infestation levels were not statistically tested. Only data from the years with the most abundant collections were considered for the

calculation of the infestation levels, i.e. 2014 for Italy and Switzerland, and 2015 for the Netherlands.

## Results

Fruits from a total of 165 plant species were collected in the three countries, providing 24,165 *D. suzukii* adults, 4153 in Italy, 15,527 in the Netherlands and 4485 in Switzerland. The list of the plant species from which *D. suzukii* emerged is provided in Table 1, with quantitative information on the sampling and emergence. The plant species from which no *D. suzukii* emerged are listed in Table 2. In total, 84 plant species from 19 families gave rise to adult emergence, 39 species in Italy, 52 in the Netherlands and 24 in Switzerland. Forty-two of these are recorded for the first time as hosts of *D. suzukii* in the field, of which six had already been found to be suitable for larval development in laboratory studies (Baroffio et al. 2014; Lee et al. 2015; Poyet et al. 2015) (Table 1). Thirty-eight host species are not native to any of the three investigated countries. Fifty are commonly found in the wild in at least one of the three regions, 53 are commonly planted as ornamental and 16 are commonly cultivated fruits.

The rate of occurrence is presented for all fruit species collected in at least five different sites  $\times$  year (Fig. 1). *Drosophila suzukii* emerged from fruit of *Frangula alnus*, *Sambucus nigra*, *Rubus fruticosus* aggr., *Rubus caesius*, *Prunus laurocerasus* and *Phytolacca americana* in at least 80 % of the sites  $\times$  years. In contrast, *Sorbus aucuparia*, *Crataegus monogyna*, *Rosa canina*, *Mahonia aquifolium*, *Prunus cerasifera*, *Paris quadrifolia* and *Viburnum lantana* were only occasional hosts, with *D. suzukii* emerging at maximum 20 % of the sites  $\times$  years. Among the 81 plant species that did not provide *D. suzukii*, only six were frequently collected (at least five sites  $\times$  year) and 10 of them had been found as field or laboratory hosts in previous studies (Table 2).

The infestation levels are presented for Italy 2014, Switzerland (Ticino only) 2014 and the Netherlands 2015 in Figs. 2, 3 and 4. In Italy, the highest levels, measured as the number of flies emerging per dm<sup>2</sup> of fruit, were found in plants of the genera *Sambucus* and *Rubus* followed by *Frangula alnus* (Fig. 2). The high score in *Sambucus ebulus*, a species sampled only in Italy, was obtained on the basis of a single site providing an enormous amount of flies emerging from its small fruits. In Switzerland, the same parameter provided slightly different results, with the highest scores obtained from a few figs (*Ficus carica*), followed by *Frangula alnus*, *Phytolacca americana* and *Prunus padus*. *Rubus* spp. showed a similar infestation level in Italy and Switzerland, while fewer flies emerged

from *Sambucus* spp. in Switzerland. In the Netherlands, where the infestation level was measured as the number of *D. suzukii* adults per kg of fruit, the highest scores were also obtained by *Rubus* spp. but, more surprisingly, also by *Cornus sanguinea*, a species that was not or only poorly attacked in Switzerland and Italy. Similarly, *Taxus baccata* was heavily infested in the Netherlands and much less so in the two other countries. Other heavily attacked species in the Netherlands included three *Vaccinium* species and several species that also scored high in the other countries such as *Prunus laurocerasus*, *Sambucus nigra* and *Frangula alnus* (Fig. 4).

The fruits that were found early in the season, i.e. before June 1 in Italy, before July 1 in the Netherlands and on June 6 in Ticino (no fruits were found in Ticino at the first survey on May 9) are listed in Table 3. In general, fruits found in spring fall into two categories: those that are produced in spring and those that are produced in late summer and autumn but that last until spring of the following year. Fruit species of the first category were all infested by *D. suzukii*. In contrast, most fruit species that are able to last over winter were not infested and, from the six species that were, five of them contained *D. suzukii* only in autumn (Table 3). Only *Cotoneaster lacteus* fruits were found infested in November and again throughout April. A large collection of fruits was carried out in the Netherlands on 4 December 2014 to assess whether some could potentially host overwintering larvae. However, no fly emerged, even from fruits that had provided numerous flies until October. Plant species that were sampled that date included *Crataegus monogyna*, *Prunus spinosa*, *Rosa canina*, *Rosa rugosa*, *Rubus fruticosus* aggr., *Symphoricarpos albus* and *Taxus baccata*.

## Discussion

This survey confirmed that *D. suzukii* is highly polyphagous and can attack and develop in a wide range of fruits of wild and ornamental plants as well as cultivated fruits. Forty-one plant species, both indigenous and exotic, have been added to the list of suitable hosts. Several hosts such as *Rubus* spp., *Sambucus* spp., *Prunus* spp., *Lonicera* spp. and *Frangula alnus* were consistently found throughout the sites and years as being heavily infested, confirming similar observations made in other studies (Mitsui et al. 2010; Baroffio et al. 2014; Lee et al. 2015; Poyet et al. 2015). Other results were more surprising. In particular, we did not expect so many adults emerging from species such as *Rosa* spp. and *Malus baccata*, which tend to have a rather tough skin. Similarly, flies were obtained from many other plant species considered to be unsuitable in laboratory tests carried out by Poyet et al. (2015),

**Table 1** Fruit species from which *D. suzukii* adults emerged, in Italy (IT), the Netherlands (NL) and Switzerland (CH: Ticino TI; Jura JU), in 2014 and 2015

Species (family)	New host record <sup>a</sup>	Main habitat/ purpose <sup>b</sup>	Native/ Exotic <sup>c</sup>	IT 2014	IT 2015	NL 2014	NL 2015	CH-TI 2014	CH-JU 2014
<i>Actinidia chinensis</i> Planch. (Actinidiaceae)		F	E	1/1					
<i>Amelanchier lamarckii</i> F.G. Schr. (Rosaceae)	✓	O/W	E			0/2	1/2		
<i>Amelanchier ovalis</i> Medik. (Rosaceae)	✓	W/O	N	1/2					
<i>Arbutus unedo</i> L. (Ericaceae)		W/O	N	1/2	0/1				
<i>Arum italicum</i> Mill. (Araceae)	✓	O/W	N	0/1		0/2	1/1		
<i>Cornus alba</i> L. (Cornaceae)	✓	O	E				1/1		
<i>Cornus kousa</i> Hance (Cornaceae)		O	E				1/1		
<i>Cornus mas</i> L (Cornaceae)		W/O	N	1/3	1/1		1/1	1/1	
<i>Cornus sanguinea</i> L. (Cornaceae)		W/O	N	2/4	0/1	2/4	2/2	0/6	0/3
<i>Cotoneaster franchetii</i> Boiss. (Rosaceae)	✓	O	E				1/1		
<i>Cotoneaster lacteus</i> W.W. Smith (Rosaceae)		O	E	1/2	0/1				
<i>Cotoneaster rehderi</i> Pojark. (Rosaceae)	✓	O	E				1/1		
<i>Crataegus chrysoarpa</i> Ashe (Rosaceae)	✓	O	E				1/1		
<i>Crataegus monogyna</i> Jacq. (Rosaceae)	✓	W/O	N	0/4	0/1	2/6		0/1	0/2
<i>Daphne mezereum</i> L. (Thymelaeaceae)	✓	W	N	1/2	0/1				
<i>Duchesnea indica</i> (Andr.) Focke (Rosaceae)	Lab	O/W	E	1/2		0/1	3/3	2/5	
<i>Eriobotrya japonica</i> (Thunb.) Lindl. (Rosac.)		O	E	1/2					
<i>Ficus carica</i> (L.) (Moraceae)		F	N	1/3				3/3	
<i>Fragaria vesca</i> L. (Rosaceae)		W	N	0/1	1/1		1/1	9/22	
<i>Frangula alnus</i> Mill. (Rhamnaceae)		W	N	3/3		2/2	1/1	3/3	
<i>Gaultheria x wisleyensis</i> M.&M. (Ericaceae)	✓	O	E				1/1		
<i>Hippophae rhamnoides</i> L. (Elaeagnaceae)		W/F	N	0/1				1/1	
<i>Lonicera alpigena</i> L. (Caprifoliaceae)		W	N	2/3	2/3				
<i>Lonicera caerulea</i> L. (Caprifoliaceae)		W	N	1/1	1/1				
<i>Lonicera caprifolium</i> L. (Caprifoliaceae)	✓	W/O	N		0/1		2/2		
<i>Lonicera ferdinandii</i> Franch. (Caprifoliaceae)	✓	O	E				1/1		
<i>Lonicera nigra</i> L. (Caprifoliaceae)		W	N	1/2	2/2				
<i>Lonicera nitida</i> E. H. Wilson (Caprifoliaceae)	Lab	O	E				2/2		
<i>Lonicera</i> sp (Caprifoliaceae)								6/8	
<i>Lonicera xylosteum</i> L. (Caprifoliaceae)		W	N	1/3	1/4				
<i>Lycium barbarum</i> L. (Solanaceae)	✓	O/F/W	N	1/2					
<i>Mahonia aquifolium</i> (Pursh) Nutt. (Berberid.)	Lab	O	E			0/1	1/5		
<i>Mahonia</i> sp. (Berberidaceae)		O	E	0/1	1/1				
<i>Malus baccata</i> Borkh. (Rosaceae)	✓	O	E				1/1		
<i>Paris quadrifolia</i> L. (Melanthiaceae)	✓	O	N	0/3	0/1			1/1	
<i>Parthenocissus quinquefolia</i> (L.) (Vitaceae)		O	E	0/2				2/2	
<i>Photinia beauverdiana</i> C. K. Schn. (Rosaceae)	✓	O	E				1/1		
<i>Photinia villosa</i> (Thunb.) DC. (Rosaceae)	✓	O	E				1/1		
<i>Photinia prunifolia</i> Lindl. (Rosaceae)	✓	O	E				1/1		
<i>Phytolacca americana</i> L. (Phytolaccaceae)		O/W	E	4/4	0/1	0/1		5/5	
<i>Phytolacca esculenta</i> Van Houtte (Phytolacc.)	✓	O/W	E				1/1		
<i>Polygonatum multiflorum</i> (L.) All. (Liliaceae)	✓	W	N				1/1	0/1	
<i>Prunus armeniaca</i> L. (Rosaceae)		F	E	1/1					
<i>Prunus avium</i> (L.) (Rosaceae)		W/F/O	N			0/2	1/2	5/10	
<i>Prunus cerasifera</i> Ehrh. (Rosaceae)		O/W	E	1/1					
<i>Prunus cerasus</i> L. (Rosaceae)		F/W	N	1/1	0/1				



**Table 1** continued

Species (family)	New host record <sup>a</sup>	Main habitat/ purpose <sup>b</sup>	Native/ Exotic <sup>c</sup>	IT 2014	IT 2015	NL 2014	NL 2015	CH-TI 2014	CH-JU 2014
<i>Prunus domestica</i> L. (Rosaceae)		F	E					2/2	0/1
<i>Prunus laurocerasus</i> L. (Rosaceae)		O/W	E	1/1	0/1	1/1	1/1	2/2	
<i>Prunus lusitanica</i> L. (Rosaceae)		O	E	1/1					
<i>Prunus mahaleb</i> L. (Rosaceae)		W/O	N	2/3	1/3				
<i>Prunus padus</i> L. (Rosaceae)	Lab	W/O	N			0/1	1/1	1/1	
<i>Prunus serotina</i> Ehrhart (Rosaceae)		W	E			1/2	1/1		
<i>Prunus spinosa</i> L. (Rosaceae)	Lab	W/O	N	1/5	0/2	2/5	2/3		0/1
<i>Pyracantha</i> sp. (Rosaceae)		O	E				1/1		
<i>Rhamnus cathartica</i> L. (Rhamnaceae)		W	N				2/2		
<i>Ribes rubrum</i> L. (Rosaceae)	Lab	F	N			0/1	2/3	0/1	
<i>Rosa acicularis</i> Lindl. (Rosaceae)	✓	O	E				1/1		
<i>Rosa canina</i> L. (Rosaceae)	✓	W/O	N	0/7	0/5	3/5	0/2		
<i>Rosa glauca</i> Pourr. (Rosaceae)	✓	O/W	N				1/1		
<i>Rosa pimpinellifolia</i> L. (Rosaceae)	✓	O/W	N				1/1		
<i>Rosa rugosa</i> Thunb. (Rosaceae)	✓	W/O	E			0/1	3/3		
<i>Rubus caesius</i> L. (Rosaceae)	✓	W	N		1/2		3/3		
<i>Rubus fruticosus</i> aggr. (Rosaceae)		W/F	N	4/5		8/9	6/6	29/32 <sup>d</sup>	5/7
<i>Rubus idaeus</i> L. (Rosaceae)		W/F	N	2/2		0/3	1/2	12/16	
<i>Rubus phoenicolasius</i> Maxim. (Rosaceae)	✓	F	E					2/2	
<i>Rubus saxatilis</i> L. (Rosaceae)	✓	W	N		2/2			0/1	
<i>Sambucus ebulus</i> L. (Adoxaceae)		W	N	1/2					
<i>Sambucus nigra</i> L. (Adoxaceae)		W	N	2/3	0/2	33/34	4/4	5/8	2/3
<i>Sambucus racemosa</i> L. (Adoxaceae)		W/O	N	1/3	5/6	0/3	1/1	4/5	
<i>Solanum dulcamara</i> L. (Solanaceae)		W	N	0/3	0/1	0/2	3/4	1/6	
<i>Solanum nigrum</i> L. (Solanaceae)		W	N	0/4			1/4	1/1	
<i>Sorbus aria</i> (L.) (Rosaceae)	✓	W/O	N	1/3	0/1				
<i>Sorbus aucuparia</i> L. (Rosaceae)	✓	W/O	N	0/4	0/1	1/4	0/1		
<i>Symphoricarpos albus</i> (L.) (Caprifoliaceae)		O/W	E	0/1		2/3	0/1	0/1	
<i>Tamus communis</i> L. (Dioscoreaceae)	✓	W	N	2/4				2/3	
<i>Taxus baccata</i> L. (Taxaceae)		O/W	N	2/3		0/1	1/1	1/1	
<i>Vaccinium myrtilloides</i> Michx. (Ericaceae)	✓	F/O	E				1/1		
<i>Vaccinium myrtillus</i> L. (Ericaceae)		W/F	N	1/1				1/1	
<i>Vaccinium oldhamii</i> Miquel. (Ericaceae)	✓	O/F	E				1/1		
<i>Vaccinium praestans</i> Lamb. (Ericaceae)	✓	O	E				1/1		
<i>Vaccinium vitis-idea</i> L. (Ericaceae)		O	E	0/1			1/1		
<i>Viburnum lantana</i> L. (Adoxaceae)		W/O	N	0/1	1/3	0/1			
<i>Viburnum rhytidophyllum</i> Hemsl. (Adoxaceae)	✓	O	E				1/1		
<i>Vitis vinifera</i> L. (Vitaceae)		F	N	1/1				0/3	

a/b: a number of sites where *D. suzukii* was obtained; b number of sites where the fruit was collected

<sup>a</sup> New host record: ✓ = species not yet reported in the literature as host in the field, based on Cini et al. (2012), Baroffio et al. (2014), Asplen et al. (2015) and Lee et al. 2015; Lab species not yet found as host in the field but suitable host in laboratory tests in Baroffio et al. (2014), Lee et al. (2015) or Poyet et al. (2015)

<sup>b</sup> Main habitat/purpose: W commonly found in the wild in at least one of the three regions, O commonly planted as ornamental, F commonly planted as fruit crop; minor habitats/purposes are not indicated

<sup>c</sup> Native (N) = native in at least one of the investigated regions; Exotic (E) exotic in the three regions

<sup>d</sup> In Ticino, *Rubus fruticosus* aggr. may have also included *Rubus caesius*

**Table 2** Fruit species from which no *D. suzukii* adults emerged, in Italy (IT), the Netherlands (NL) and Switzerland (CH: Ticino TI; Jura JU), in 2014 and 2015, with the number of sites sampled

Species (family)	Known host of <i>D. suzukii</i> <sup>1</sup>	Number of sites where fruits were sampled					
		IT 2014	IT 2015	NL 2014	NL 2015	CH-TI 2014	CH-JU 2014
<i>Actaea spicata</i> L. (Ranunculaceae)		2	1			3	
<i>Ampelopsis brevipedunculata</i> (Max.) Tr. (Vitaceae)			1				
<i>Aronia x prunifolia</i> (Rosaceae)		1					
<i>Asparagus acutifolius</i> L. (Asparagaceae)		1					
<i>Asparagus officinalis</i> L. (Asparagaceae)					1		
<i>Atropa bella-donna</i> L. (Solanaceae)	Lab						1
<i>Aucuba japonica</i> Thunberg (Garryaceae)	Field	1	2				
<i>Berberis x media</i> (Berberidaceae)		1					
<b><i>Berberis vulgaris</i></b> L. (Berberidaceae)		3				3	
<i>Berberis</i> sp. (Berberidaceae)					2		
<i>Bryonia dioica</i> Jacq. (Cucurbitaceae)		1	1				
<i>Callicarpa bodinieri</i> H. Lév. (Lamiaceae)		1			1		
<i>Cephalotaxus harringtonia</i> (K. ex F.) Koch (Cephalotaxaceae)		1					
<i>Chamaerops</i> sp. (Arecaceae)		1					
<i>Convallaria majalis</i> L. (Nolinoideae)		2					
<i>Cotoneaster acutifolius</i> Turcz. (Rosaceae)		1					
<i>Cotoneaster dammeri</i> C. K. Schneid. (Rosaceae)		1					
<i>Cotoneaster horizontalis</i> Decne. (Rosaceae)	Field	1			1	1	
<i>Cotoneaster microphyllus</i> Wall. ex Lindl. (Rosaceae)		2					
<i>Cotoneaster salicifolius</i> Franch. (Rosaceae)		1	2				
<i>Cotoneaster suecicus</i> G.Klotz (Rosaceae)					1		
<i>Cotoneaster</i> × <i>watereri</i> Exell (Rosaceae)				1			
<i>Crataegus azarolus</i> L. (Rosaceae)			1				
<i>Crataegus coccinea</i> L. (Rosaceae)		1					
<i>Crataegus crus-galli</i> L. (Rosaceae)		1					
<i>Crataegus kansuensis</i> E. H. Wilson (Rosaceae)					1		
<i>Crataegus laevigata</i> (Poir.) DC. (Rosaceae)		1	1				
<i>Crataegus</i> sp. (Rosaceae)		2					
<i>Diospyros kaki</i> Thunberg (Ebenaceae)	Field	1					
<i>Euonymus europaeus</i> L. (Celastraceae)		1		3			
<i>Gaultheria shallon</i> Pursh (Ericaceae)					1		
<i>Gaultheria</i> sp. 1 (Ericaceae)		1					
<i>Gaultheria</i> sp. 2 (Ericaceae)		1					
<b><i>Hedera helix</i></b> L. (Araliaceae)		1	3	1	3		
<i>Hypericum</i> sp. (Hypericaceae)					1		
<i>Hypericum androsaemum</i> L. (Hypericaceae)		1	1				
<i>Ilex aquifolium</i> L. (Aquifoliaceae)		1		1	1		
<i>Ilex</i> sp. (Aquifoliaceae)		1					
<i>Juniperus</i> sp. (Cupressaceae)		1					
<i>Laurus nobilis</i> L. (Lauraceae)		1	1				
<i>Ligustrum lucidum</i> W. T. Aiton (Oleaceae)		1					
<b><i>Ligustrum vulgare</i></b> L. (Oleaceae)		4	1	2	1	2	
<i>Lonicera etrusca</i> Santi (Caprifoliaceae)		1					
<i>Lonicera henryi</i> Hemsl. (Caprifoliaceae)		1			1		
<i>Lonicera periclymenum</i> L. (Caprifoliaceae)	Field			1			
<i>Lonicera pileata</i> Oliv. (Caprifoliaceae)		1					



**Table 2** continued

Species (family)	Known host of <i>D. sukuzii</i> <sup>1</sup>	Number of sites where fruits were sampled					
		IT 2014	IT 2015	NL 2014	NL 2015	CH-TI 2014	CH-JU 2014
<i>Malus floribunda</i> Siebold ex Van Houtte (Rosaceae)		1	1				
<i>Malus</i> x Red Sentinel (Rosaceae)					1		
<i>Mespilus germanica</i> L. (Rosaceae)		1				1	
<i>Morus alba</i> L. (Moraceae)	Field	1					
<i>Myrteola</i> sp. (Myrtaceae)		1					
<i>Myrtus communis</i> L. (Myrtaceae)			2				
<i>Nandina domestica</i> Thunb. (Berberidaceae)		2	2				
<i>Olea europaea</i> L. cv. Leccino (Oleaceae)		1					
<i>Parthenocissus tricuspidata</i> (Sieb. & Zucc.) Planch. (Vitaceae)		1					
<i>Phillyrea angustifolia</i> L. (Oleaceae)			2				
<i>Phillyrea latifolia</i> L. (Oleaceae)			3				
<i>Prunus persica</i> (L.) var. florepleno (Rosaceae)	Field		1				
<i>Punica granatum</i> L. (Lythraceae)		2					
<i>Pyracantha coccinea</i> M. Roem. (Rosaceae)		1			1		
<i>Pyracantha 'navaho'</i> (Rosaceae)		1	1				
<i>Rhamnus pumila</i> Turra (Rhamnaceae)		1	1				
<i>Rhodotypos scandens</i> (Thunb.) Makino (Rosaceae)		1					
<i>Ribes alpinum</i> L. (Rosaceae)	Field			1			
<i>Rosa pendulina</i> L. (Rosaceae)		2					
<i>Rubus ulmifolius</i> Schott. (Rosaceae)			4				
<b><i>Ruscus aculeatus</i> L.</b> (Asparagaceae)		3	5				
<i>Skimmia japonica</i> Thunberg (Rutaceae)					1		
<i>Smilax aspera</i> L. (Smilacaceae)			1				
<i>Solanum pseudocapsicum</i> L. (Solanaceae)		1					
<i>Solanum sisymbriifolium</i> Lam. (Solanaceae)		1					
<i>Sorbus chamaemespilus</i> (L.) Crantz (Rosaceae)		2	1				
<i>Sorbus intermedia</i> (Ehrh.) Pers. (Rosaceae)				1			
<i>Symphoricarpos x chenaultii</i> Hanc. (Caprifoliaceae)					1		
<i>Symphoricarpos orbiculatus</i> Moen. (Caprifoliaceae)					1		
<i>Vaccinium uliginosum</i> L. (Ericaceae)		1					
<b><i>Viburnum opulus</i> L.</b> (Adoxaceae)	Field	2		4		2	2
<i>Viburnum tinus</i> L. (Adoxaceae)		1					
<i>Viscum album</i> L. (Santalaceae)	Lab	1					
<i>Vitis labrusca</i> L. (Vitaceae)		2					
<i>Withania somnifera</i> (L.) Dunal, (Solanaceae)			1				

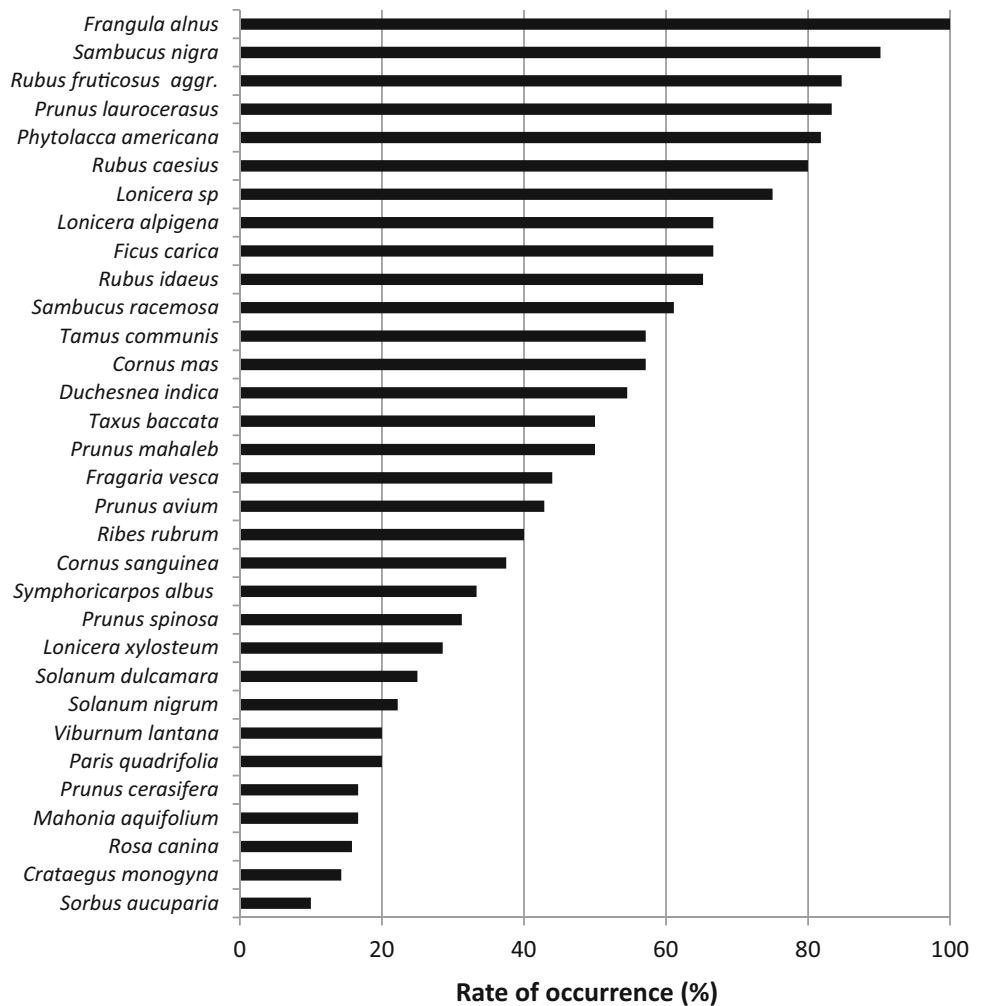
In bold: six species collected in at least five combinations of sites × years

<sup>1</sup> Known host of *D. sukuzii*: Field = plant already recorded as host in the field, based on the reviews of Cini et al. (2012), Baroffio et al. (2014) and Lee et al. (2015); Lab species not yet found as host in the field but suitable host in laboratory tests in Baroffio et al. (2014), Lee et al. (2015) or Poyet et al. (2015)

such as *Sorbus aria*, *Sorbus aucuparia*, *Polygonatum multiflorum*, *Paris quadrifolia* and *Crataegus monogyna*. These unexpected infestations can be partly explained by the very high population levels of *D. sukuzii* in the second half of 2014 (Italy and Switzerland) and 2015 (the Netherlands). Moreover, although surveys focused on ripe, undamaged fruits, it is likely that some adults emerged

from “hard” fruits, such as *Malus baccata*, resulted from eggs laid in unnoticed damaged fruits. Lee et al. (2015) also obtained *D. sukuzii* from field-collected fruits that appeared unsuitable in laboratory tests. They attributed these discrepancies to differences in fruit suitability among picked (laboratory) versus hanging (field) fruit and the timing of sampling.

**Fig. 1** Rate of occurrence of *D. suzukii* in the host plants in which it emerged, expressed as the % of sites × years in which *D. suzukii* was found. Only the fruits found in at least 5 sites × years are presented

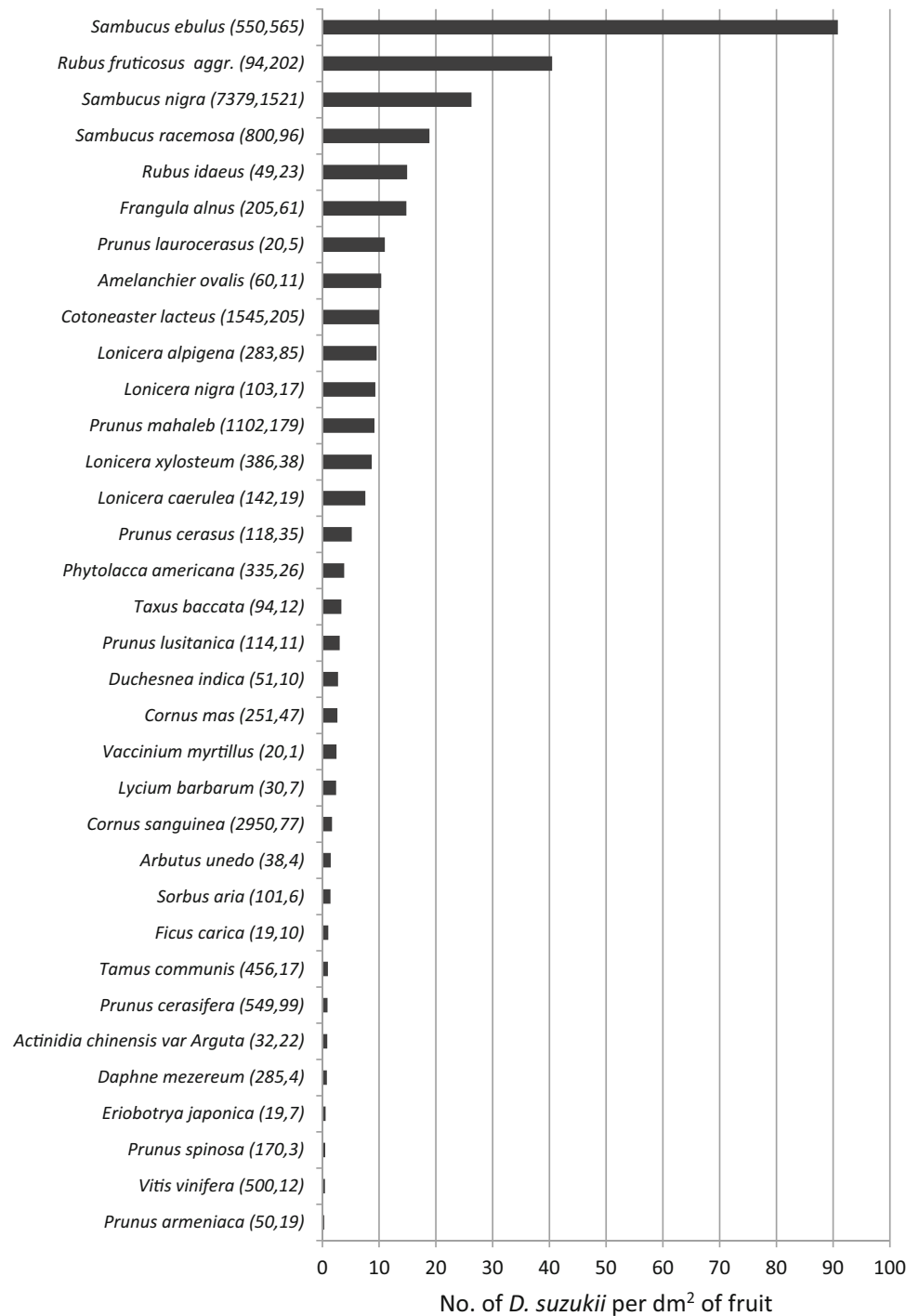


Eighty-one plant species did not give rise to *D. suzukii* adults, among which ten have been reported as hosts in other field surveys or laboratory tests (Baroffio et al. 2014; Lee et al. 2015; Poyet et al. 2015). These negative results must be considered with great caution. Most of the fruit species from which nothing emerged were collected in low numbers or at few sites. Laboratory tests could be carried out to confirm the unsuitability of these fruits, taking into account that discrepancies between laboratory tests and field surveys may occur (Lee et al. 2015). Only the six fruit species collected in high numbers in at least five combinations of sites × years can be regarded as “unsuitable”: *Actaea spicata*, *Berberis vulgaris*, *Hedera helix*, *Ligustrum vulgare*, *Ruscus aculeatus* and *Viburnum opulus*, even though, for the latter species, some larval development but no adult emergence had been observed in laboratory tests (Poyet et al. 2015).

This survey also illustrated the close association between *D. suzukii* and invasive plants. Forty plants identified as hosts are exotic to the survey areas and many of them are considered as invasive species. The interaction

between *D. suzukii* and the invasive American black cherry *Prunus serotina* has been studied in France by Poyet et al. (2014), who suggest that the heavy infestation of *Prunus serotina* fruits by *D. suzukii* could reduce the life span of fruits and their attractiveness to seed consumers and dispersers. In contrast, *Prunus serotina* could represent a suitable plant reservoir enhancing *D. suzukii* invasion. A similar scenario was proposed by Asplen et al. (2015) regarding the invasion of the European buckthorn, *Rhamnus cathartica*, in North America, which was found to be a suitable host of *D. suzukii* both in North America and in our study. The invasive ‘Himalayan’ blackberry, *Rubus armeniacus*, is also known to favour the spread and abundance of *D. suzukii* in berry production systems in Western North America (Klick et al. 2016). Besides *Prunus serotina*, other important invasive plants infested by *D. suzukii* in our samples include, e.g. *Duchesnea indica*, *Phytolacca americana* and *Phytolacca esculenta*, *Prunus laurocerasus*, *Rosa rugosa* and *Symphoricarpos albus*. The interactions between the invasion processes of *D. suzukii* and these invasive plants should be further investigated.

**Fig. 2** No. of *D. suzukii* emerged per dm<sup>2</sup> of fruit collected in 2014 in Italy. Numbers in parentheses after the fruits' names indicate the total number of fruits collected and the total number of *D. suzukii* adults emerged from these fruits, respectively. Only fruits from which *D. suzukii* emerged are shown

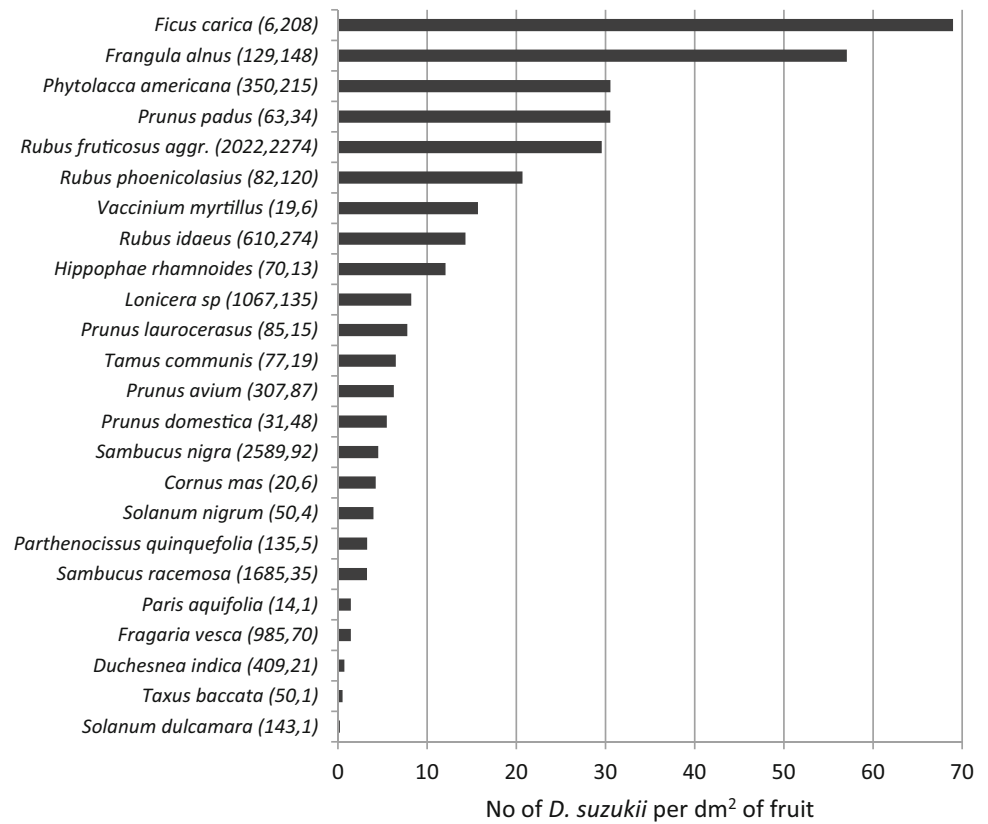


### Implications for sustainable *Drosophila suzukii* management

Knowing the realised host range and the preferred host plants outside cultivated habitats is essential for the development of sustainable IPM strategies against *D.*

*suzukii* (Klick et al. 2016; Lee et al. 2015). Pelton et al. (2016) showed that the amount of woodland in the landscape positively affects early season crop risk and the high numbers of *D. suzukii* in the woods have implications for understanding overwintering. Non-crop hosts in the vicinity of susceptible fruit crops may also enhance *D. suzukii*

**Fig. 3** No. of *D. suzukii* emerged per dm<sup>2</sup> of fruit collected in 2014 in Ticino, Switzerland. Numbers in parentheses after the fruits' names indicate the total number of fruits collected and the total number of *D. suzukii* adults emerged from these fruits, respectively. Only fruits from which *D. suzukii* emerged are shown

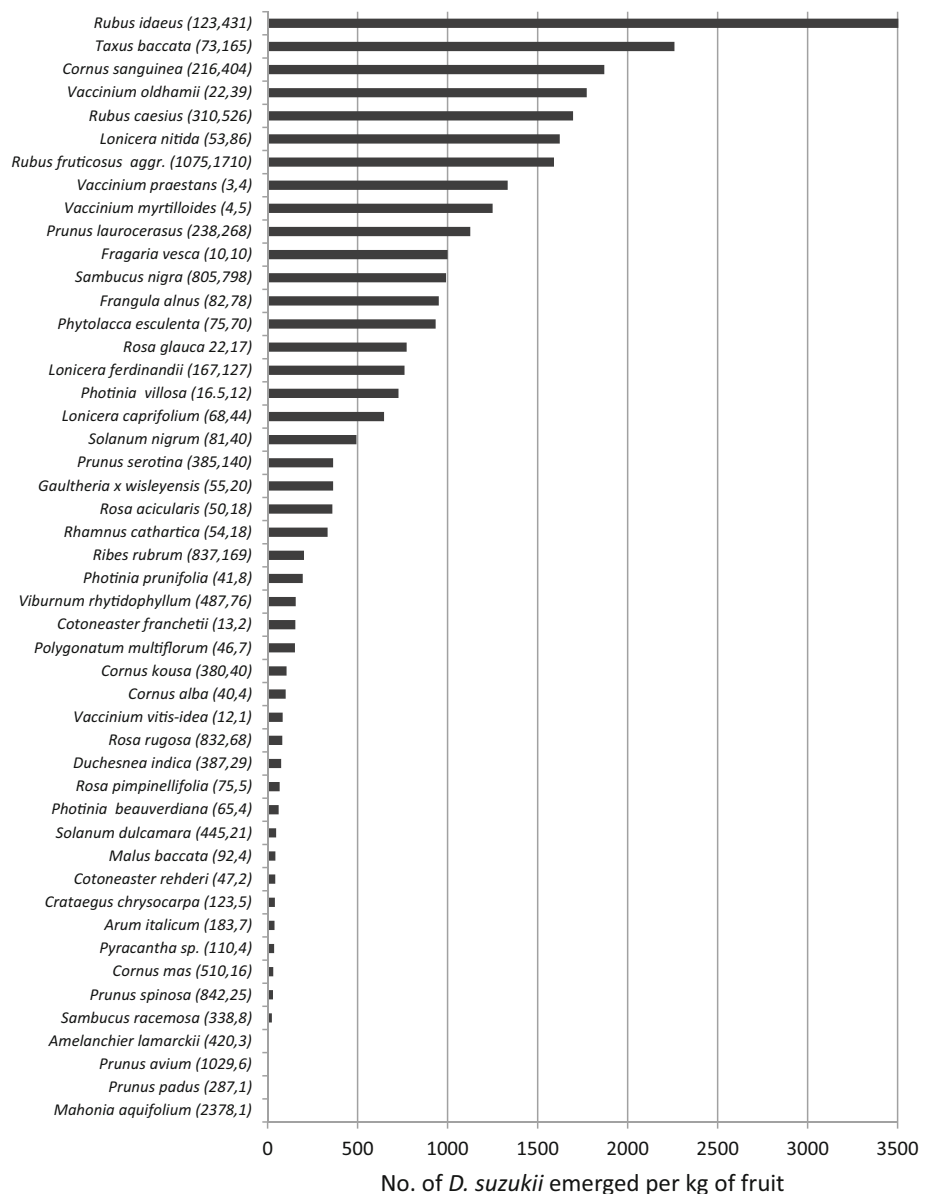


populations before or during the crop season, as shown by Klick et al. (2016) in a raspberry crop system in Western North America. These alternative hosts may also be used as refuges for *D. suzukii* when crops are sprayed with insecticides. Therefore the management of these non-crop hosts should be integrated in control strategies. For example, our results and other host range studies (Lee et al. 2015; Poyet et al. 2015) now allow us to advise on suitable and unsuitable ornamental plants to be used in the vicinity of susceptible crops. Species such as *Cornus* spp., *Lonicera* spp., *Prunus* spp., *Sambucus* spp. and *Taxus baccata*, which are abundantly used as hedge plants in Europe, should be avoided. In contrast, there is now sufficient evidence showing that, e.g. *Ligustrum* spp., *Viburnum* spp., *Crataegus* spp. or *Pyracantha* spp. do not increase populations of *D. suzukii* on site. Similarly, field margins could be cleared of susceptible wild plants (Klick et al. 2016). However, the management of wild hosts in the surroundings of crops is often more problematic than ornamental hosts because of the high number of highly susceptible species and the difficulty in managing them in areas that do not always belong to the fruit producer. Furthermore, more should be known on the natural dispersal capacities of *D. suzukii* to determine the areas requiring management. If dispersal studies show that *D. suzukii* can be attracted over

long distances, removing native wild host plants from a large area may become unpractical and have a negative effect on the functioning of local ecosystems.

The fruiting period of host plants is also an essential consideration when developing management strategies. In Europe, populations of *D. suzukii* often dramatically increase from spring to autumn, due to the high number of generations (Asplen et al. 2015). Only a few plants produce fruits in spring, suggesting that the availability of suitable fruits in spring is a key element in the population dynamics of *D. suzukii*. Therefore, efforts should be made to control the presence of these early fruits in the surroundings of fruit crops and orchards (Asplen et al. 2015; Poyet et al. 2015). Not only non-crop hosts should be controlled. In the surveyed area in Northern Italy and the Netherlands, the first heavy infestations occur on abandoned or untreated cherry trees, which probably play an important role in the local increase of *D. suzukii* populations in summer (Ioriatti et al. 2015; Helsen and van der Sluis unpublished data). This survey showed that all plants fruiting in spring were attacked by *D. suzukii*. In contrast, most fruits that are formed in autumn and overwinter until spring were used neither as overwintering hosts nor as early hosts in spring, with the possible exception of fruits of *C. lacteus* that were found attacked in April. We did not

**Fig. 4** No. of *D. suzukii* emerged per kg of fruit collected in 2015 in the Netherlands. Numbers in parentheses after the fruits' names indicate the total weight of fruits collected and the total number of *D. suzukii* adults emerged from these fruits, respectively. Only fruits from which *D. suzukii* emerged are shown



find any evidence that *D. suzukii* larvae may overwinter in fruits. In Northern Italy, the monitoring and dissection of female flies throughout the winter showed that *D. suzukii* overwinters as adults in reproductive diapause from November to April (Zerulla et al. 2015).

More generally, the ability of *D. suzukii* to attack such a large number of widely distributed ornamental and wild host plants strongly suggests the need for an area-wide control approach. Since insecticides are often not effective (e.g. Rogers et al. 2016) and cannot be used in many of the non-crop habitats, in particular forests, and since sanitation is impossible on a large scale, classical biological control through the introduction of specific parasitoids from the

region of origin of the fly could be a long term solution (Haye et al. 2016; Daane et al. 2016). Preliminary studies in Japan have suggested that some larval parasitoids are specific to *D. suzukii* (Kasuya et al. 2013; Nomano et al. 2015) and recent surveys in East Asia have shown that larval parasitism rates are not negligible, e.g. less than 10 % in the Tokyo region (Kasuya et al. 2013), up to 16 % in South Korea (Daane et al. 2016) and over 50 % in Yunnan, China (Kenis unpublished data). It would be worth assessing the suitability of these parasitoids for introduction into invaded areas, including the evaluation of potential non-target effects of such introductions on the community of native Drosophilidae.

**Table 3** Host suitability of fruits found early in the year: before June 1st in Italy, before July 1st in the Netherlands and on June 6 in Switzerland

Fruits formed in spring		Fruits formed in summer or autumn and lasting until the following year		
Suitable hosts	Host suitability not proved in this study	Suitable hosts in autumn and spring	Suitable host only in autumn	Host suitability not proved in this study
<i>Amelanchier lamarckii</i>		<i>Cotoneaster lacteus</i>	<i>Crataegus monogyna</i> <sup>a</sup>	<i>Aucuba japonica</i> <sup>b</sup>
<i>Eriobotrya japonica</i>			<i>Duchesnea indica</i> <sup>a</sup>	<i>Cotoneaster microphyllus</i>
<i>Fragaria vesca</i>			<i>Prunus laurocerasus</i> <sup>a</sup>	<i>Cotoneaster salicifolia</i>
<i>Lonicera xylosteum</i>			<i>Pyracantha</i> sp.	<i>Hedera helix</i>
<i>Mahonia aquifolium</i>			<i>Rosa canina</i>	<i>Ilex aquifolium</i>
<i>Prunus avium</i>				<i>Laurus nobilis</i>
<i>Prunus mahaleb</i>				<i>Ligustrum vulgare</i>
<i>Prunus padus</i>				<i>Malus floribunda</i>
<i>Ribes rubrum</i>				<i>Mespilus germanica</i>
<i>Rubus</i> sp.				<i>Morus alba</i> <sup>b</sup>
<i>Sambucus racemosa</i>				<i>Nandina domestica</i>
				<i>Pyracantha</i> ‘navaho’
				<i>Ruscus aculeatus</i>
				<i>Viburnum opulus</i>

<sup>a</sup> For these species, only few fruits pass the winter

<sup>b</sup> Plants already recorded as suitable hosts in the field in Japan (Lee et al. 2015)

## Author contribution statement

MK conceived the experiment and coordinated the writing up of the manuscript, which was carried out jointly by all authors. All authors participated in the field sampling and laboratory rearing.

**Acknowledgments** We thank Andrea Battisti and three anonymous reviewers for their useful comments on the manuscript. The research leading to these results has received funding from the European Union’s Seventh Framework programme for research, technological development and demonstration under grant agreement number 613678 (DROPSA).

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