



Natural Host Plant Survey of the Economically Important Fruit Flies (Diptera: Tephritidae) of Chiapas, Mexico

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NATURAL HOST PLANT SURVEY OF THE ECONOMICALLY IMPORTANT FRUIT FLIES (DIPTERA: TEPHRITIDAE) OF CHIAPAS, MEXICO

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ABSTRACT

We provide a natural host-plant list of some of the fruit flies (Diptera: Tephritidae) reported for the State of Chiapas. Out of 92 plant species sampled, 39 species representing 12 plant families were identified as natural host of *Anastrepha distincta*, *A. fraterculus*, *A. leptozona*, *A. ludens*, *A. obliqua*, *A. serpentina*, *A. striata* and *Toxotrypana curvicauda*.

RESUMEN

Se presenta una lista de las plantas que han sido identificadas como hospederas naturales de algunas de las especies de moscas de la fruta (Diptera: Tephritidae) reportadas en el estado de Chiapas, México. De 92 especies vegetales muestreadas, 39 especies, pertenecientes a 12 familias fueron identificadas como hospederas naturales de *Anastrepha distincta*, *A. fraterculus*, *A. leptozona*, *A. ludens*, *A. obliqua*, *A. serpentina*, *A. striata* y *Toxotrypana curvicauda*.

Fruit flies (Diptera: Tephritidae) are well known in Mexico for the devastating impact they can have on commercial fruit production. Entire fruit growing regions have been forced out of business due to heavy infestations of these widely distributed insects (Aluja and Liedo 1986). Larvae-infested fruit usually drop from the tree and rot, rendering them useless for human consumption. The presence of these insects also elicits neighboring regions and countries to apply severe quarantine restrictions to prevent their introduction to fruit growing regions free of fruit flies. Even though economic damage is basically restricted to semicommercial and commercial orchards and isolated backyard trees, fruit flies infest a wide variety of wild native host plants and even small populations serve as reservoirs from which commercial fruit orchards are invaded. Aluja and Liedo (1986) proposed a series of ecologically based management practices such as habitat manipulation, trap crops and orchard designs, where emphasis is placed on trying to stop the continuous flow of populations from one host to another. They nevertheless noted that one prerequisite that must be fulfilled before even attempting to experimentally achieve this goal, is the complete identification of all cultivated and wild host plants for each of the fruit fly species causing economic damage.

We define a natural host as a plant bearing fruit or any other tissue where female fruit flies lay eggs in nature and the emerging larvae are able to survive and reach the adult stage with varying degrees of fitness. Some host plant records in the literature are misleading because they are based on oviposition induced under artificial conditions on fruits presented to flies which had been deprived of an oviposition substrate for days. Such practices have led researchers to report fruits such as prickly pear (*Opuntia* sp.), banana (*Musa* sp.), squash (*Cucurbita mexicana*), walnut (*Juglans regia*), papaya (*Carica papaya*) and garden bean (*Phaseolus vulgaris*) as hosts of *Anastrepha ludens* (Baker et al. 1944, Ebeling 1950 and Weems 1963). Such reports are sometimes used in quarantine protocols to limit the movement of these and other fruits from one country to another, an unfortunate fact since it is almost impossible for these plants to serve as hosts under natural conditions.

Some host plant records for fruit flies found in Mexico are commonly cited in the literature but, unfortunately, many are very old and have not been confirmed since their first publication (Arriaga-Martinez 1979, Baker et al. 1944, Bush 1957, 1962, Coronado 1964, Greene 1934, McPhail and Berry 1936, Plummer et al., 1941, Ramos 1975, Shaw 1947, Stone 1942 and Weems 1963). We suspect that some host records may be the result of misidentifications of both plant and fly species. Ballou (1945), Blanchard (1937), Briceño (1975), Caraballo (1981), Cuculiza and Torres (1975), Eskafi and Cunningham (1987), Fernandez-Yepez (1953), Guaglioni (1966), Herrera and Viñas (1977), Jirón and Zeledón (1979), Korytkowski and Ojeda-Peña (1968), Lima (1934), Lutz and Lima (1918), McAlister (1936), Malavasi et al. (1980), Martorell (1939), Norrbom (1982), Olarte (1980), Rosillo (1953), Wasbauer (1972) and Whervin (1974) also published records for hosts that although not necessarily found in Mexico are nevertheless attacked by fruit fly species occurring in that country. Norrbom (1985) provides an excellent summary of most of the published host records for all *Anastrepha* species.

In this paper, we provide an updated list of the natural host plants of some of the fruit flies identified by Aluja et al. (1987) in the State of Chiapas.

MATERIALS AND METHODS

The study area comprised the Soconusco Region and the Mazapa Valley in the State of Chiapas, Mexico. A detailed description of these areas is given in Aluja et al. (1987).

Fruits from known and potential host plants were collected in commercial and semicommercial orchards, backyard gardens and areas covered with wild native vegetation from March 1982 to December 1985. Individual samples consisted of fallen fruit (80%) and fruit still on the tree (20%) and ranged in number from 1 to 120 fruits depending on the availability of a particular species or variety. The degree of ripeness varied from very green to very ripe. Samples were individually labeled, placed in plastic bags and transported to the laboratory in well cushioned and aerated styrofoam boxes to avoid mortality of immatures due to overheating and physical contact.

In the laboratory each bag was weighed and the number of fruits per sample recorded. Fruits were then immersed for 5 seconds in a 10% solution of sodium benzoate and water (to inhibit growth of fungi and decomposing bacteria) and placed in "maturation chambers". These are 18 by 33 by 25 cm styrofoam boxes, adapted to hold a wire screen basket on which fruits were placed (Fig. 1). The bottom of each box was covered with vermiculite or sawdust to provide an adequate pupation substrate for the third instar larvae dropping from infested fruit. The cover of the "maturation chamber" had a 10 by 28 cm screen that permitted air to flow freely but stopped any emerging fruit flies or fruit fly parasitoids (see Aluja 1985 for details). It is important to note that when collecting fruit from the tree, care should be taken to prevent the fruit from falling to the ground. If the fruit ruptures, high larval mortality may be caused by the sodium benzoate solution. Figure 2 shows a tool that is very useful for the collection of fruit from tall branches.

Depending on the ripeness or decomposition stage of the fruit samples, these were kept in the "maturation chambers" for a period of 4 to 14 days. Periodic inspections and dissections of randomly selected samples determined when most of the larvae had left the fruit. For each dissection we recorded the number and stage of remaining live or

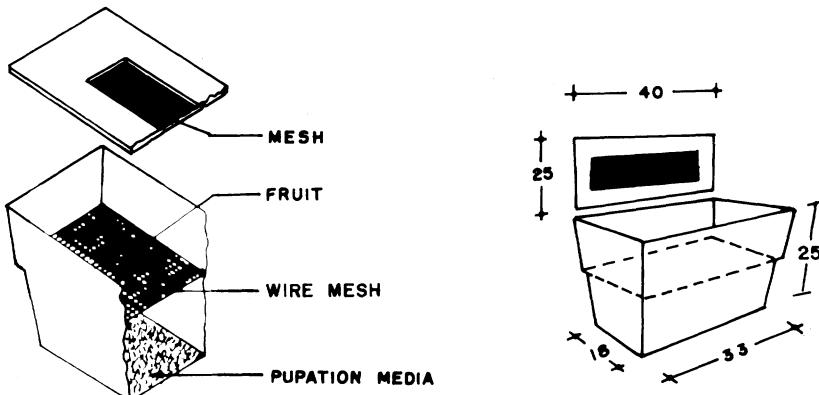


Fig. 1. "Maturation Chamber" for the collection of fruit fly larvae and pupae from infested fruit (dimensions are in cm).

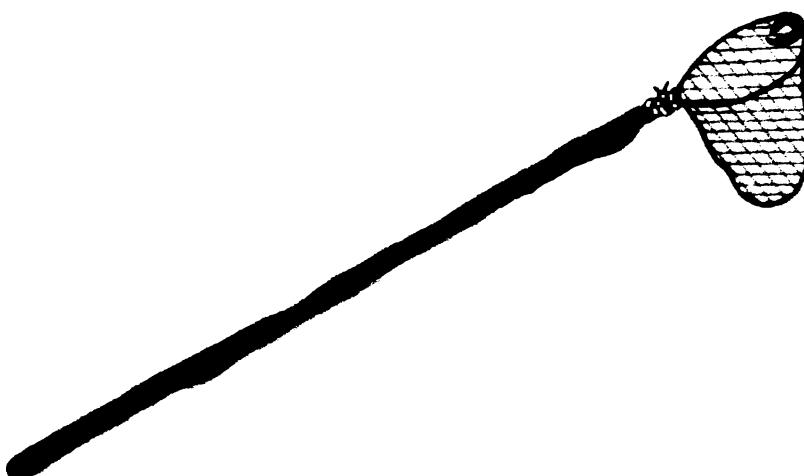


Fig. 2. Fruit collection net with sharp hook for cutting of thick pedicels.

dead larvae. At the same time the sawdust was carefully inspected and sieved to collect all pupae and larvae, which were placed in 500 ml plastic containers half filled with moist soil. Containers were covered with a fine screen capable of allowing aeration but stopping the emerging fruit flies and parasitoids from escaping. Water was regularly added to the soil in order to maintain optimal humidity conditions. The laboratory in which the plastic containers were kept had an average temperature of $28 \pm 3^\circ\text{C}$ and RH of $55 \pm 5\%$.

The containers were inspected daily and any fruit fly or parasitoid that had emerged was removed. Part of this biological material was killed and identified and part used for demographic studies. The identification process is described in Aluja et al. (1987). Host plant identifications were based on Tejada (1980), Miranda (1976) and corroborated by Mario Cabrera (Universidad Autonoma de Chiapas).

RESULTS

A total of 28,830 samples representing 92 species from 24 plant families were taken (see Table 1). Of these, 39 species from 12 families turned out to be natural hosts of the fruit fly genera *Anastrepha* and *Toxotrypana* (see Table 2). The number of hosts identified for each fruit fly species ranged from 1 to 14: *A. ludens* (14), *A. serpentina* (11), *A. obliqua* (11), *A. fraterculus* (10), *A. distincta* (4), *A. leptozona* (2), *A. striata* (1) and *Toxotrypana curvicauda* (1).

DISCUSSION AND CONCLUSIONS

Our results increase by 10 the number of natural hosts reported for the fruit flies of Chiapas (Aluja 1985 reports 24 natural hosts).

It is important to note that the information provided here refers only to the positive identification of a plant species as a natural host of fruit flies. It does not indicate the degree of infestation nor the relative frequency of infestation. Aluja and Liedo (1986) have shown that simply recording a plant as a host provides little information since some hosts may be only occasionally exploited. Usually it is observed that a few fruit

TABLE 1. PLANT SPECIES SAMPLED IN THE SOCONUSCO REGION AND THE MAZAPA DE MADERO VALLEY (CHIAPAS, MEXICO) TO DETERMINE THEIR STATUS AS HOST PLANTS OF THE LOCAL FRUIT FLIES.

Spanish	Common name	English	Scientific name
Aceituno		Wild olive	<i>Simarouba glauca</i> DC.
Aguacate		Avocado	<i>Persea americana</i> Mill.
Algodoncillo		?	?
Almendra		Tropical almond	<i>Terminalia catappa</i> L.
Anona		Sweetsop	<i>Annona squamosa</i> L.
Anona colorada		Custard apple	<i>A. reticulata</i> L.
Baricoco		?	<i>Micropholis mexicana</i> Gilly
Cachofla		?	?
Caco		Coco plum	<i>Chrysobalanus icaco</i> L.
Café		Coffee	<i>Coffea arabica</i> L.
Caimito		Star apple	<i>Chrysophyllum cainito</i> L.
Capulín		Strawberry tree	<i>Muntingia calabura</i> L.
Caspirol		Inga	<i>Inga lauriana</i> Willd.
Chachalaco		?	?
Chalum		Inga	<i>Inga micheliana</i> Harms
Chayote		Chayote	<i>Sechium edule</i> (Jacq.) Swartz
Chelel		Inga	<i>Inga leptoloba</i> Schl.
Chico zapote		Sapodilla	<i>Manilkara achras</i> Mill.
Chileamate		?	?
Chilindrón		?	?
Chincuya		?	<i>Annona purpurea</i> Mets.
Cidra		Citron	<i>Citrus medica</i> L.
Coroso		Corozo palm fruit	<i>Orbignya cohune</i> (Mart.)
Cuajilote		Cuachilote	<i>Paermenteria edulis</i> D. C.
Cuajinicuil		Inga	<i>Inga spuria</i> H. et B.
Cuernavaca		?	<i>Solanum macranthum</i> Dunal.
Cupapé		?	<i>Cordia dodecandra</i> De.
Encino (zapotillo)		—	<i>Comepia polyandra</i> Rose
Granada		Pomegranate	<i>Punica granatum</i> L.
Granadilla		Passion fruit	<i>Passiflora edulis</i> Sims
Grosella		?	?
Guamuchil		?	<i>Pithecellobium dulce</i> Benth
Guanábana		Soursop	<i>Annona muricata</i> L.
Guaya		Spanish lime	<i>Melicocca bijuga</i> L.
Guayaba		Guava	<i>Psidium guajava</i> L.
Higo		Fig	<i>Ficus carica</i> L.
Huevo de mono		Inga	<i>Inga</i> sp.
Hule silvestre		?	?
Jaboncillo		Soapberry	<i>Sapindus saponaria</i> L.
Jocote		Purple mombin	<i>Spondias purpurea</i> L.
Jobo de pava		Mombin	<i>Spondias mombin</i> L.
Jushte		Breadnut	<i>Brosimum alicastrum</i> Sw.
Lechón		?	?
Lima chiche		?	<i>Citrus</i> sp.
Lima limón		Sweet lime	<i>C. limetta</i> Rissso (Christm.)
Lombriero		?	?
Maluco		?	?
Mandarina		Tangerine	<i>Citrus deliciosa</i> Tencre
Mandarina china		Chinese tangerine	<i>Citrus</i> sp.
Mandarina criolla		Creol tangerine	<i>C. reticulata</i> Blco.

TABLE 1. (Continued)

Common name		
Spanish	English	Scientific name
Mango	Mango	<i>Mangifera indica</i> L.
Manzanilla	Hawthorn	<i>Crataegus</i> sp.
Mapahuite	?	<i>Trichilia</i> sp.
Marachán	?	?
Marañón	Cashew nut	<i>Anacardium occidentale</i> L.
Matapalo	?	<i>Ficus tecolotensis</i> Standley
Matasano	White sapote	<i>Casimiroa edulis</i> Llave & Lex.
Matzú	?	<i>Cordia alba</i> Roem & Schult.
Melocotón	Peach	<i>Prunus persica</i> (L.) Batsch
Melón	Musk melon	<i>Cucumis melo</i> L.
Nance	Nanche	<i>Byrsonima crassifolia</i> (L.) HBK
Naranja agria	Sour orange	<i>Citrus aurantium</i> L.
Naranja dulce	Orange	<i>C. sinensis</i> (L.) Osbeck
Níspero	Loquat	<i>Eriobotrya japonica</i> Lindl.
Pacaya	Pacaya palm fruit	<i>Chamaedorea aguilariana</i> Standley & Steyermark
Pan de la India	Jack fruit tree	<i>Artocarpus heterophyllus</i> Lam.
Pan de maría	?	?
Pan de palo	Breadfruit	<i>Artocarpus altilis</i> (Perkins) Fosb.
Papause	Illama	<i>Annona diversifolia</i> Saff.
Papaya comercial	Papaya	<i>Carica papaya</i> L.
Papaya silvestre	Wild papaya	<i>Carica caudiflora</i> JMacq.
Patashte	?	<i>Theobroma bicolor</i> Humb. & Bonpl.
Paterna	Inga	<i>Inga paterna</i> Harms
Pepino silvestre	Wild cucumber	<i>Cucumis</i> sp.
Pera de agua	?	<i>Syzygium malaceensis</i> L.
Piñón	?	<i>Jatropha curcas</i> L.
Pitanga	Surinam cherry	<i>Syzygium uniflora</i> L.
Pomarrosa	Rose apple	<i>Syzygium jambos</i> L.
Pomelo	Pummelo	<i>Citrus maxima</i> (Burm.) Merrill
Pozól	?	<i>Alchornea latifolia</i> Sw.
Quishtán	?	<i>Solanum wendlandii</i> Hook.
Sangre de toro	?	?
Sikajal	?	?
Taculchacha	?	?
Tempisque	?	<i>Mastichodendron capiri</i> var. <i>tempisque</i> (A. DC.) Cronq.
Tomate silvestre	Wild Tomato	<i>Lycopersicon</i> sp.
Toronja	Grapefurit	<i>Citrus paradisi</i> Macfadyen
Toronjil	?	<i>Rheedia edulis</i> Planch & Triana
Tuna	Prickly pear	<i>Opuntia tuna</i> (L.) Mill.
Zapote amarillo	Canistal	<i>Pouteria campechiana</i> Baehni
Zapote colorado	Mammee apple	<i>Calocarpum sapota</i> Merr.
Zapote de agua	?	<i>Pachira aquatica</i> Aubl.
Zapote negro	Black sapote	<i>Diospyros ebenaster</i> Retz.
Zunzapote	?	<i>Licania platypus</i> Fritsch

TABLE 2. NATURAL HOST PLANTS OF SOME OF THE FRUIT FLIES OF CHIAPAS, MEXICO. LISTED IN ORDER OF ECONOMIC IMPORTANCE.

Fruit fly species	Common local name of host plant	Scientific name of host plant
<i>Anastrepha ludens</i>	Mango	<i>Mangifera indica</i> L.
	Naranja dulce	<i>Citrus sinensis</i> L. (Osbeck)
	Naranja agria	<i>C. aurantium</i> L.
	Pomelo	<i>C. maxima</i> (Burm.) Merrill
	Mandarina criolla	<i>C. reticulata</i> Blco.
	Toronja	<i>C. paradisi</i> Macfady
	Tangerina	<i>C. deliciosa</i> Tenore
	Cidra	<i>C. medica</i> L.
	Lima limón	<i>C. limetta</i> Risso (Christm.)
	Mandarina China	<i>C. sp.</i>
	Naranja injerto	<i>C. sp.</i>
	Matasano	<i>Casimiroa edulis</i> Llave & Lex.
	Anona	<i>Annona squamosa</i> L.
	Tempisque	<i>Mastichodendron capiri</i> var. <i>tempisque</i> (A. D. C.) Cronq.
<i>Anastrepha serpentina</i>	Zapote colorado	<i>Calocarpum sapota</i> Merr.
	Chico zapote	<i>Manilkara achras</i> (Mill.)
	Caimito	<i>Chrysophyllum cainito</i> L.
	Zapote amarillo	<i>Pouteria campechiana</i> Baehni
	Baricoco	<i>Micropholis mexicana</i> L.
	Pomelo	<i>Citrus maxima</i> (Burm.) Merrill
	Naranja agria	<i>C. aurantium</i> L.
	Naranja dulce	<i>C. sinensis</i> (L.) Osbeck
	Pozol	<i>Alchornea latifolia</i> Sw.
	Nance	<i>Byssonia crassifolia</i> (L.) HBK
	Mango	<i>Mangifera indica</i> L.
<i>Anastrepha obliqua</i>	Mango	<i>Mangifera indica</i> L.
	Guayaba	<i>Psidium guajava</i> L.
	Pomarrosa	<i>Syzygium jambos</i> L.
	Pera China	<i>S. malaceensis</i> L.
	Jobo de pava	<i>Spondias mombin</i> L.
	Jocote	<i>S. purpurea</i> L.
	Jushte	<i>Brosimum alicastrum</i> (Ramon)
	Nispero	<i>Eriobotrya japonica</i> Lindl.
	Pozol	<i>Alchornea latifolia</i> Sw.
	Grosella	?
	Manzanilla	<i>Crataegus</i> sp.
<i>Anastrepha fraterculus</i>	Guayaba	<i>Psidium guajava</i> L.
	Pomarrosa	<i>Syzygium jambos</i> L.
	Pozol	<i>Alchornea latifolia</i> Sw.
	Pitanga	<i>Syzygium uniflora</i> L.
	Tempisque	<i>Mastichodendron capiri</i> var. <i>tempisque</i> (A. D. C.) Cronq.
	Almendra	<i>Terminalia catappa</i> L.
	Cafe	<i>Coffea arabica</i> L.
	Mango	<i>Mangifera indica</i> L.
	Grosella	?
	Manzanilla	<i>Crataegus</i> sp.
<i>Anastrepha distincta</i>	Chalum	<i>Inga michelianiana</i> Harms

TABLE 2. (Continued)

Fruit fly species	Common local name of host plant	Scientific name of host plant
<i>Anastrepha leptozona</i>	Caspirol	<i>I. lauriana</i> Willd.
	Cuajinicuil	<i>I. spuria</i> H. et B.
	Paterna	<i>I. paterna</i> Harms
<i>Anastrepha striata</i>	Baricoco	<i>Micropholis mexicana</i> Gilly
	Manzanilla	<i>Crataegus</i> sp.
<i>Toxotrypana curvicauda</i>	Guayaba	<i>Psidium guajava</i> L.
	Papaya	<i>Carica papaya</i> L.

species are preferred in any given region. We therefore suggest that any future host records also provide complete information as to how important the host is and what the host utilization patterns are in nature (Aluja et al. unpublished information).

After carefully studying the host plant records reported in the literature, we have concluded that the range of our sampling schemes need to be increased to consider more species of families that at first would not appear to be an adequate substrate for larval development. For example, the families Euphorbiaceae, Passifloraceae and Ebenaceae are very common in the Soconusco Region, but were only sampled in low numbers. An extended sampling effort may soon allow us to identify the natural hosts of *A. alveata*, *A. balloui*, *A. bicolor*, *A. chilcayae*, *A. montei*, *A. pallens* and *A. tripunctata*, the other fruit flies identified in the State of Chiapas by Aluja et al. (1987).

We consider that some of the hosts reported in the literature may be the result of misidentifications or sampling errors. It seems unlikely that *Manihot esculenta* is a host of *A. obliqua* as Guaglumi (1966) suggests. Caraballo (1981) reports *Theobroma* sp. as a host of *A. fraterculus* which is a phenomenon not observed in the varieties of *Theobroma* sp. planted in Mexico. Most of the hosts reported by Korytkowski & Ojeda-Peña (1968) had never been reported before for the species they indicate.

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