

## IDENTIFICATION AND CHARACTERIZATION OF CARMOVIRUS ON CARNATION (*Dianthus caryophyllus* L.) IN WEST JAVA, INDONESIA

Erniawati Diningsih<sup>1)</sup>, Gede Suastika<sup>2\*)</sup>, Tri Asmira Damayanti<sup>2)</sup> and Slamet Susanto<sup>3)</sup>

<sup>1)</sup> Indonesian Ornamental Crop Research Institute (IOCRI)

Jl. Raya Ciherang PO BOX 8 SDL 43253 Cianjur West Java Indonesia

<sup>2)</sup> Department of Plant Protection, Faculty of Agriculture, Bogor Agricultural University,  
Campus IPB Darmaga, Jl. Kamper, Bogor 16680 West Java Indonesia

<sup>3)</sup> Department of Agronomy and Horticulture, Faculty of Agriculture, Bogor Agricultural University,  
Campus IPB Darmaga, Jl. Meranti, Bogor 16680 West Java Indonesia

<sup>\*)</sup> Corresponding author E-mail: gsuast@gmail.com

**Received: February 9, 2015/ Accepted: May 13, 2015**

### ABSTRACT

Carnation has a highly economic demand of cut flower in Indonesia. Field observations in West Java Indonesia was conducted in order to find the typical mottle symptoms that was a suspect caused by a virus disease. Identification of the virus was respectively conducted by performing ELISA test with four anti sera and characterizations held by bioassay, observing of virion particles, detecting of nucleic acid by RT-PCR and nucleotide sequencing. Total of 403 samples were collected from plants with or no virus-like symptoms. Among those all tested, 83% were found to be infected by *Carnation mottle virus* (CarMV), but negatively against *Carnation ringspot virus* (CRSV), *Carnation latent virus* (CLV), and *Carnation vein mottle virus* (CVMV) antisera. By mechanical inoculation, the virus was able to infect systemically *Cenopodium quinoa* and locally infect on others. However on *Phalaenopsis* sp and *Gomprena globosa*, there was symptomless found. The isometric CarMV particles size was approximately 30 nm. RT-PCR using specific primers of CP gene of CarMV successfully amplified a DNA sized 1000 bp. CarMV West Java Indonesian (Idn-WJ) isolates possessed the highest nucleotide and amino acid homology with CarMV from Spain and was in the same cluster with CarMV from China, Taiwan and Israel.

Keyword: *Dianthus caryophyllus*, RT-PCR, sequence, Tombusviridae

### INTRODUCTION

Carnation (*Dianthus caryophyllus*) is one of the major cut flowers in the world since the flower is beautiful with many variation of colors, shapes, and types. The flowers are produced worldwide including Western Europe, Eastern Europe, Latin America, United State of America, Central America, Japan, South east Asia, Australia, Israel (Lisa, 1995), India (Singh *et al.*, 2005) and Turkey (Cevik *et al.*, 2010).

In Indonesia cut flower production of carnation is lower than other ornamental plants i.e. orchid, rose, and chrysanthemum. It is mostly due to lack of pathogen free stocks. Recently, mottle was disease found in various carnation plantations in West Java. Virus-like infected plants showed mild to severe light green to yellowish stripes with unclear boundaries, dark green leaf vein, and curly leaf edges (Figure 1). Occasionally dwarf and curly shoot symptoms were found on certain carnation cultivars.

Some viruses reported naturally infected carnation in the field that were *Carnation ringspot virus* (CRSV), *Carnation latent virus* (CLV), *Carnation vein mottle virus* (CVMV), and *Carnation mottle virus* (CarMV) (Lisa, 1995; Sanchez-Navarro *et al.*, 1999; Fauquet *et al.*, 2005). Some of these viruses cause similar symptoms. In some carnation production countries, these viruses were stated as the most important virus affecting carnation plants (Singh *et al.*, 2005; Ahmed *et al.*, 2012). Since phenotype symptoms is difficult in determining

**Accredited SK No.: 81/DIKTI/Kep/2011**

**<http://dx.doi.org/10.17503/Agrivita-2015-37-2-p108-114>**

Erniawati Diningsih *et al.*: Identification and Characterization of Carmovirus on Carnation.....

the virus causal of the disease, thus here we report the identification and characterization the causal of mottle diseases on carnation in West Java, Indonesia.

## MATERIALS AND METHODS

The nucleotide sequence reported in this paper is available in the DDBJ/EMBL/GenBank databases under the accession number KP119182.

### Time and Place

Research was conducted at the Laboratory of Virology and greenhouse of Indonesia Ornamental Crop Research Institute (IOCRI), and Laboratory of Virology, Department of Plant Protection, Faculty of Agriculture, Bogor Agricultural University.

### Samples Collection

Samples were collected from five carnation production areas at Cipanas, Ciputri (Pacet District, Cianjur Regency), Cihideung, Ciwangun (Parongpong District, West Bandung Regency), and Pangalengan (Bandung Regency) in West Java, Indonesia. Randomly, 50-100 samples were collected from each area which consisted of 25-50 samples symptomatic and 25-50 asymptomatic. A total of 403 symptomatic and asymptomatic sample plants were collected and analysed.

### Serological Detection

The diseases incidence of viruses were determined serologically (ELISA test) using four antisera such as CarMV antiserum (Agdia, USA), CRSV antiserum (Agdia, USA), CLV antiserum (Agdia, USA), and CVMV antiserum (DSMZ, Germany). ELISA tests were conducted according to protocol provided by Agdia and DSMZ.

### Host Range Test

To characterize the biological properties of carnation main virus, host range test was conducted using CarMV of West Java (Idn-WJ) isolate. Inocula were prepared by propagating virus in *C. quinoa* and grinding the *C. quinoa* infected leave in phosphate buffer 0.025 M pH 7.6 (Sing *et al.*, 2005). Sap of *C. quinoa* infected leave was mechanically inoculated by finger to upper of leave of test plants. The sap was inoculated on nine plant species from seven families i.e. Amaranaceae (*Gomphrena globosa*), Solanaceae (*Lycopersicon esculentum*, *Nicotiana clevelandii*), Cucurbitaceae (*Cucumis sativus*), Begoniaceae (*Begonia* sp), Compositae (*Ageratum conyzoides*), Chenopodiaceae (*Chenopodium quinoa* and *C. amaranticolor*), and Orchidaceae (*Phalaenopsis* sp). All tested plants were maintained in insect free green house. The incubation period, disease incidence, and type of symptom were observed daily for at least up to 30 days post inoculation. ELISA test was conducted on test plants at the end of observation.



Figure 1. Typical mottle symptoms on infected carnation plants (a) introducing cultivar (White Candy) and (b) local clone

### Electron Microscopy

Morphological observation of virus particles was performed using negative staining of transmission electron microscopy (TEM) in PT Eijkman, Jakarta, Indonesia. Virus particles from infected leaves were prepared as described by Harris (1999).

### RT-PCR

Total RNA was extracted from infected leaf tissues of carnation using commercial kit RNeasy plant mini kit (Qiagen, Germany). For reverse transcription reaction (10 µl), the cDNA was synthesized from total RNA using Moloney Murine Leukemia Virus (MMuLV) (Thermo) at 25°C for 5 min, 42°C for 60 min and 70°C for 15 min. Amplification of the cDNA was carried out using a pair of primer specific to CP gene of CarMV. The forward primer was BC57 (5'-GATCGCGATGAATCCCACTGTGC -3') and reverse primer was BC58 (5'-TCACATCCTA TAACAACCATG-3') with expected PCR product size 1000 bp (Cevik *et al.*, 2010) and PCR program according to Raikhy *et al.* (2006).

The PCR product was directly sequenced. The nucleotide sequences of the CP gene were aligned with those of other CarMV from GenBank database using Clustal-W (Thompson *et al.*, 1994). Sequences homology analysis of CarMV CP gene were performed using Bio Edit version 7.05 (<http://www.mbio.ncsu.edu/BioEdit/bioedit.html>), and phylogenetic tree was constructed using MEGA 5.0 software with the neighbor-joining algorithm and 1,000 bootstrap replications (Tamura *et al.*, 2011).

## RESULTS AND DISCUSSION

### Symptoms on Infected Carnation

Most of the carnation plants grown in the field showed light-dark green/mottle symptoms on their leaves (Figure 1). The symptoms varied depended on the carnation cultivars planted in each carnation cultivation area. Chlorosis/mottle and malformation of leaves were dominantly observed on infected plants in Ciputri village,

while severe light-dark spot symptom observed on infected leaves in Cipanas village. Chlorosis streak and stunting was observed dominantly in Ciwangun village. The mottle with no variation symptom on infected plants were observed in Cihideung and Pengalengan villages.

### The Disease Incidences of Viruses

Based on ELISA test, it showed that most of samples positively reacted against CarMV antiserum, and negatively against CRSV, CVMV and CLV antisera. Among all of 403 leaf samples tested, 336 samples (83%) were infected by CarMV. It was indicated that CarMV was a dominant virus on carnations in West Java-Indonesia.

### Host Range Test

Host range test on nine plant species showed that CarMV infected systemically on *C. quinoa*. However, there was no obvious symptom on *G. globosa* and *Phalaenopsis* sp. CarMV caused local infection on *Lycopersicon esculentum*, *C. amaranticolor*, *Ageratum conyzoides*, *Nicotiana clevelandii*, *Begonia* sp, and *Cucumis sativus* (Table 1).

Symptom on *L. esculentum*, *N. clevelandii*, and *C. sativus*, the local symptom was interveinal chlorosis streak, while on *C. amaranticolor* was chlorotic local lesions. Symptom on *C. quinoa* was systemic chlorotic lesions after 6-7 day post inoculation. Leaf malformation was also observed either on local or systemic infection. Rugose was observed on infected leaves of *A. conyzoides* and *Begonia* sp.

### Electron Microscopy

Electron microscopy examination of naturally infected carnation leaf sap was conducted in order to observe the isometric carmovirus like particles measuring 30 nm in diameter when stained with 2% uranyl acetat (Figure 2).

Erniawati Diningsih *et al.*: Identification and Characterization of Carmovirus on Carnation.....

Table 1. Host range test of CarMV WJ isolate

Plant family/species	Incidence	Symptoms*	ELISA
Amarantaceae			
<i>Gomphrena globosa</i>	0/15	-/-	-
Chenopodiaceae			
<i>Chenopodium quinoa</i>	15/15	CS/SM, Mf	+
<i>C. amaranticolor</i>	15/15	CLL/-	+
Compositae			
<i>Ageratum conyzoides</i>	8/10	ML/-	+
Solanaceae			
<i>Lycopersicon esculentum</i>	15/15/15	CSt, Mf/-	+
<i>Nicotiana clevelandii</i>		FLC, Mf/-	+
Begoniaceae			
<i>Begonia</i> sp	13/15	ML/-	-
Cucurbitaceae			
<i>Cucumis sativus</i>	15/15	CSt/-	+
Orchidaceae			
<i>Phalaenopsis</i> sp	0/15	-/-	-

Remarks: \* On inoculated/upper leaves; CS = Chlorotic Spot; CLL = Chlorotic Local Lesions; ML = Mottle Local; CL = Chlorosis Local; Mf = Malformation; CSt = Chlorotic stripe, FLC = Few local chlorotic, SM = Systemic mottling, - = no symptom. Inoculated/upper leaf tissues were assayed 1-2 week after inoculation. ELISA data were considered to be positive if absorbance values of were more than twice those of healthy plants.

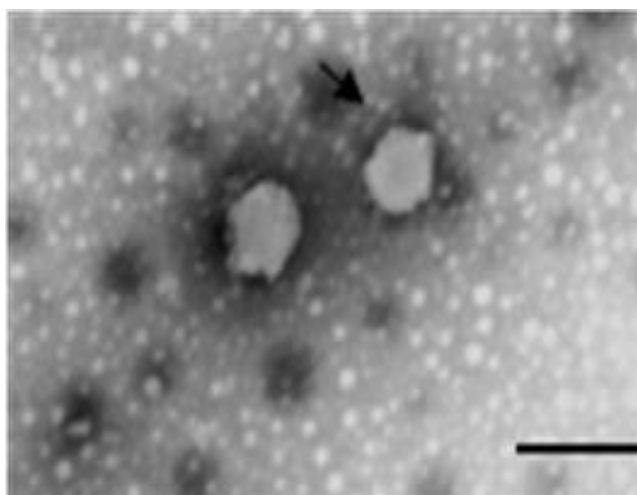


Figure 2. Transmission electron micrograph virus particles of CarMV-Idn WJ (arrow) negatively stained with 2% uranyl acetate Bar = 100 nm

### Nucleotide Sequences Analysis

The RT-PCR of all samples successfully amplified the coat protein gene of CarMV with expected size approximately 1000 bp (data not shown). Based on homology of nucleotide sequences of CP gene, all of West Java isolates were *Carnation mottle virus* (CarMV) which had similar molecular characteristic with that of isolates from the other countries. The homology of nucleotide sequences of the CP gene among isolates was ranged 92.1 to 96.6% homology for CP nucleotides sequences, and 83.5 to 95.1% homology for CP amino acid sequences (Table 2). The nucleotide sequences of the entire CP gene of WJ isolate (KP119182) had highest identity with that of CarMV from Spain (AJ309509) (97.8% identity for CP nucleotide sequences and 95.1% identity for CP amino acid sequences).

Phylogenetic tree analysis of CarMV CP gene nucleotide of WJ isolate to that of other CarMV isolates deposited in GenBank showed that CarMV WJ isolate was in the same cluster with that of isolates YN (China), Ph (Taiwan) and Isr2 (Israel), however it was distinctive to other isolates (Figure 3).

Based on the above results, it was shown that the mottle symptom on carnations in West Java, Indonesia was caused by CarMV with symptoms similarly reported previously. The typical symptoms that were mild mottle in the

young leaves and faint chlorosis in mature leaves of carnation depended on either cultivar or environmental condition (Calderón and Arbeláez, 1999).

CarMV is considered as an exotic virus since it is a quarantine pest according to quarantine regulation of The Republic of Indonesia. It might enter Indonesia via importation of vegetative propagation materials. Lacking an appropriate detection methods, no obvious virus symptoms on vegetative propagation materials, no clean culture practice in the fields and the lack of availabilities of virus free stock plants may facilitate the rapid distribution of CarMV in West Java.

The incidence of CarMV was dominant and high in the fields, indicating widely distribution of the virus in the fields as previously report in India, Turkey and California-USA (Lommel *et al.*, 1983; Singh *et al.*, 2005; Cevic *et al.*, 2010). The highly incidence of the CarMV might be facilitated by mechanical transmission via cutter such as knives or scissors during preparation of plant materials before planting or during harvesting. SCSMV was also reported rapidly distributed in Java, Indonesia facilitated with similar manner (Damayanti and Putra, 2011). CarMV can be easily transmitted by sap of diseased plants, cuttings scissors or contact between plants (Safari *et al.*, 2009).

Table 2. Homology of nucleotide (nt) and amino acid (aa) of CarMV West Java Isolates (Idn-WJ7) with those of other isolates from West Java and other country established in GenBank

Virus name	Accession number (country)	Homology	
		Nt	Aa
Isolate Idn-WJ4	KP119182 (Indonesia)	95.6	89.7
Isolate Italy-Ca4	EF622209.1 (Italy)	95.4	89.1
Isolate Netherland Ca3	EF622211.1 (Netherland)	96.6	92.1
Isolate Isr-2	AJ309501.1 (Israel)	96.6	92.4
Isolate Solan	AJ844549.1 (India)	97.2	93.6
Isolate usa-1	AJ309511.1 (USA)	96.5	92.2
Isolate MG 1669	JX207141.1 (Brazil)	97.0	93.6
Isolate sp-m	AJ309509.1 (Spain)	97.8	95.1
Isolate carEDOMEX1	KC834739.1 (Mexico)	96.8	92.7
Isolate YN	AF173879.1 (China)	97.0	92.7
Isolate Ph	HQ117872.1 (Taiwan)	96.5	91.5
Isolate FO25	EF622206.1 (New Zealand)	96.2	90.9
CPMV	M65155.1 (USA)	41.5	17.3

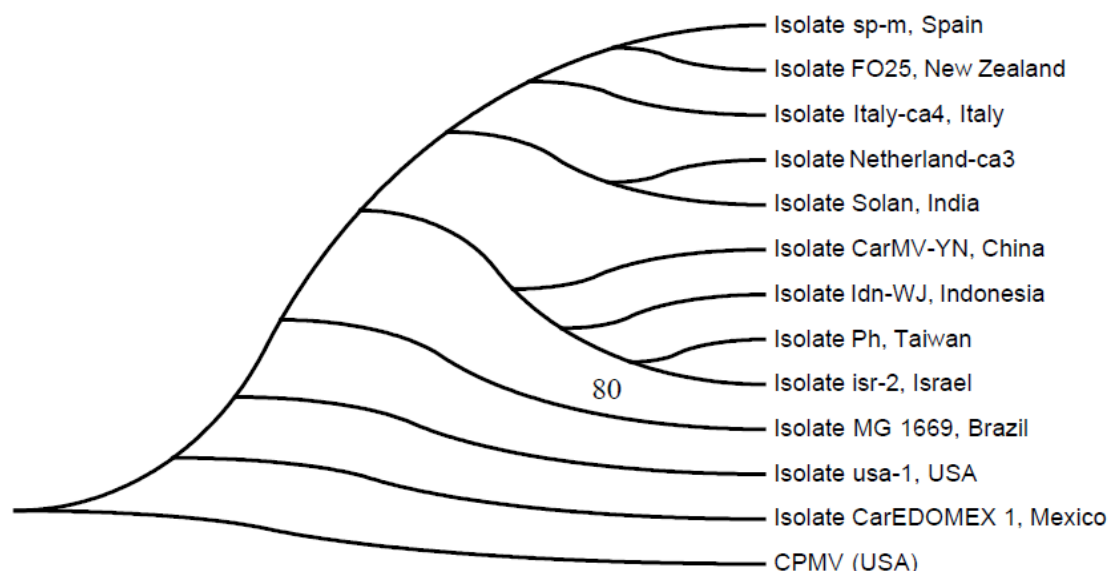


Figure 3. Phylogeny tree of the CP gene of CarMV WJ isolate. The tree was inferred by the Neighbor-Joining method using molecular evolutionary genetics analysis (MEGA) software version 5.05 based on the Clustal W alignment of CarMV sequences from Indonesia with that of 11 CarMV isolates deposited in GenBank database. The tree was rooted using *Cowpea mottle virus* (CPMV) as out groups. Bootstrap values expressed as a percentage of 1,000 replicates.

The CarMV WJ isolate might differ with that of Indian isolate (Singh *et al.*, 2005) based on its ability to infect *C. quinoa* and *G. globosa*. WJ isolate caused systemic infection on *C. quinoa* and was not able to infect *G. globosa*, while Indian isolate infects locally *C. quinoa* and *G. globosa*. The WJ isolate on *Phalaenopsis* sp was symptomless, while infected *Phalaenopsis* sp by Taiwan isolate (Zheng *et al.*, 2011) showed chlorotic ring symptom.

The different of environmental conditions between Indonesia and those of countries may cause different environmental stress which lead to different response of isolates to those tested plants. Thus, the different interactions between biotic stress (viral infection) and a abiotic stress (environmental factors) (Conti *et al.*, 2012) can cause different symptom on the same species plants (Hollings and Stone, 1964). Taken together, the WJ isolate might have differ biological characters on those isolates.

### CONCLUSION AND SUGGESTION

The results showed that CarMV was found in various varieties of carnations grown in West Java. CarMV can be transmitted mechanically on some type of test plants with different symptoms,

and the symptoms are generally localized. Except on *C. quinoa*, symptoms are lesions systemic. Isometric structure of 30 nm in diameter of virus particle was obtained, and CarMV West Java Indonesian (Idn-WJ) isolates had the highest nucleotide and amino acid homology with CarMV from Spain. Further research activities need to be done to get effective technology in the release of virus from infected plants.

### ACKNOWLEDGEMENTS

This research was supported by Agricultural Research and Development Agencies, Ministry of Agriculture, The Republic of Indonesia.

### REFERENCES

- Ahmed, A.A., E.A.H. Khatib, R.A. Dawood and A.M. Ismeil. 2012. Evaluation of tip culture and thermotherapy for elimination of Carnation latent virus and Carnation vein mottle virus from Carnation plants. *Int. J. of Virology* 8 (2): 234-239. doi: 10.3929/ijv.2012.234.239
- Calderón, O.L. and G. Arbeláez. 1999. Effect of the Carnation mottle virus on the rooting

- of the cuttings and on flower production. *Acta Hort.* 482: 179-186.
- Cevik, B., T. Bakir and G. Koca. 2010. First report of Carnation mottle virus in Turkey. *Plant Pathol.* 59: 394. <http://onlinelibrary.wiley.com/doi/10.1111/j.13653059.2009.02181.x/abstract>. Accessed August 13, 2014.
- Conti, G., M.C. Rodriguez, C.A. Manacorda and S. Asurmendi. 2012. Transgenic expression of Tobacco mosaic virus capsid and movement proteins modulate plant basal defense and biotic stress responses in *Nicotiana tabacum*. *Mol. Plant Microbe In.* 25 (10): 1370-1384.
- Damayanti, T.A. and L.K. Putra. 2011. First occurrence of sugarcane streak mosaic virus infecting sugarcane in Indonesia. *J. Gen. Plant Pathol.* 77 (1): 72-74. doi: 10.1007/s10327-010-0285-7
- Fauquet, C.M., M.A. Mayo, J. Maniloff, U. Desselberger and L.A. Ball (Eds.). 2005. Virus taxonomy: eighth report of the international committee on taxonomy of viruses. Academic Press, San Diego.
- Harris, J. Robin. 1999. Negatif staining of thinly spread biological particulates. In: Nasser Hajibagheri M.A. (Ed.), *Electron microscopy methods and protocols.* Humana press, USA. Vol. 117. p. 13-30
- Hollings, M. and O.M. Stone. 1964. Investigation of Carnation viruses. *Ann. Appl. Biol.* 53 (1): 103-118. doi: 10.1111/j.1744-7348.1964.tb03784.x
- Lisa, V. 1995. Carnation. In: Loebenstein G., Lawson R.H. and Brunt A.A. (eds.) *Virus and virus like diseases of bulb and flower crops.* p. 385-395. Wiley: New York.
- Lommel, S.A., A.H. McCain, D.E. Mayhew and T.J. Morris. 1983. Survey of commercial Carnation cultivars for four viruses in California by Indirect ELISA. *Plant Dis.* 67 (1): 53-56.
- Raikhy, G., V. Hallan, S. Kulshrestha, R. Ram and A.A. Zaidi. 2006. Multiplex PCR and genome analysis of Carnation mottle virus Indian isolate. *Curr. Sci. India* 90 (1): 74-82. <http://www.iisc.ernet.in/currsci/jan102006/74.pdf>. Accessed August 13, 2014.
- Safari, M., M.K. Habibi, G. Mosahebi and A. Dizadji. 2009. Carnation mottle virus, an important viral agent infecting Carnation cut-flower crops in Mahallat of Iran. *Commun. Agric. Appl. Biol. Sci.* 74 (3): 861-865. <http://www.ncbi.nlm.nih.gov/pubmed>. Accessed December 22, 2014.
- Sánchez-Navarro, J.A., M.C. Cañizares, E.A. Cano and V. Pallás. 1999. Simultaneous detection of five Carnation viruses by non-isotopic molecular hybridization. *J. Virol. Methods* 82 (2): 167-175. <http://www.ncbi.nlm.nih.gov/pubmed/10894633>. Accessed December 22, 2014.
- Singh, H.P., V. Hallan, G. Raikhy, S. Kulshrestha, M.L. Sharma, R. Ram, I.D. Garg and A.A. Zaidi. 2005. Characterization of an Indian isolate of Carnation mottle virus (CarMV) Infecting Carnations. *Curr. Sci. India* 88 (4): 594-601. <http://www.iisc.ernet.in/currsci/feb252005/594.pdf>. Accessed August 13, 2014.
- Tamura, K., D. Peterson, N. Peterson, G. Stecher, M. Nei and S. Kumar. 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol. Biol. Evol.* 28 (10): 2731-2739. doi: 10.1093/molbev/msr121.
- Thompson, J.D., D.G. Higgins and T.J. Gibson. 1994. CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weigh matrix choice. *Nucleic Acids Res.* 22 (22): 4673-4680.
- Zheng, Y.X., C.C. Chen and F.J. Jan. 2011. First report of Carnation mottle virus in *Phalaenopsis* Orchids. *Plant Dis.* 95 (3): 354. <http://dx.doi.org/10.1094/PDIS-10-10-0757>. Accessed August 13, 2014.